

Gemelli



Fondazione Policlinico Universitario Agostino Gemelli IRCCS
Università Cattolica del Sacro Cuore

Guido Primiano


La richiesta di competenza neurologica nel prossimo futuro

Sesta edizione

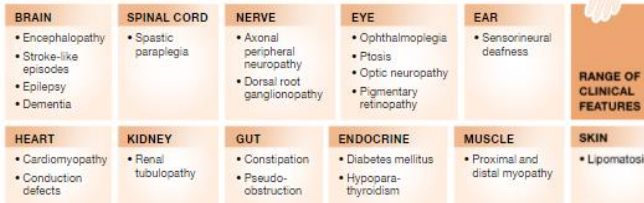
MITOCONDRIALI - Up to date

Fondazione Policlinico Universitario A. Gemelli IRCCS, Roma
Dipartimento Universitario di Neuroscienze, UCSC, Roma

Figure 1

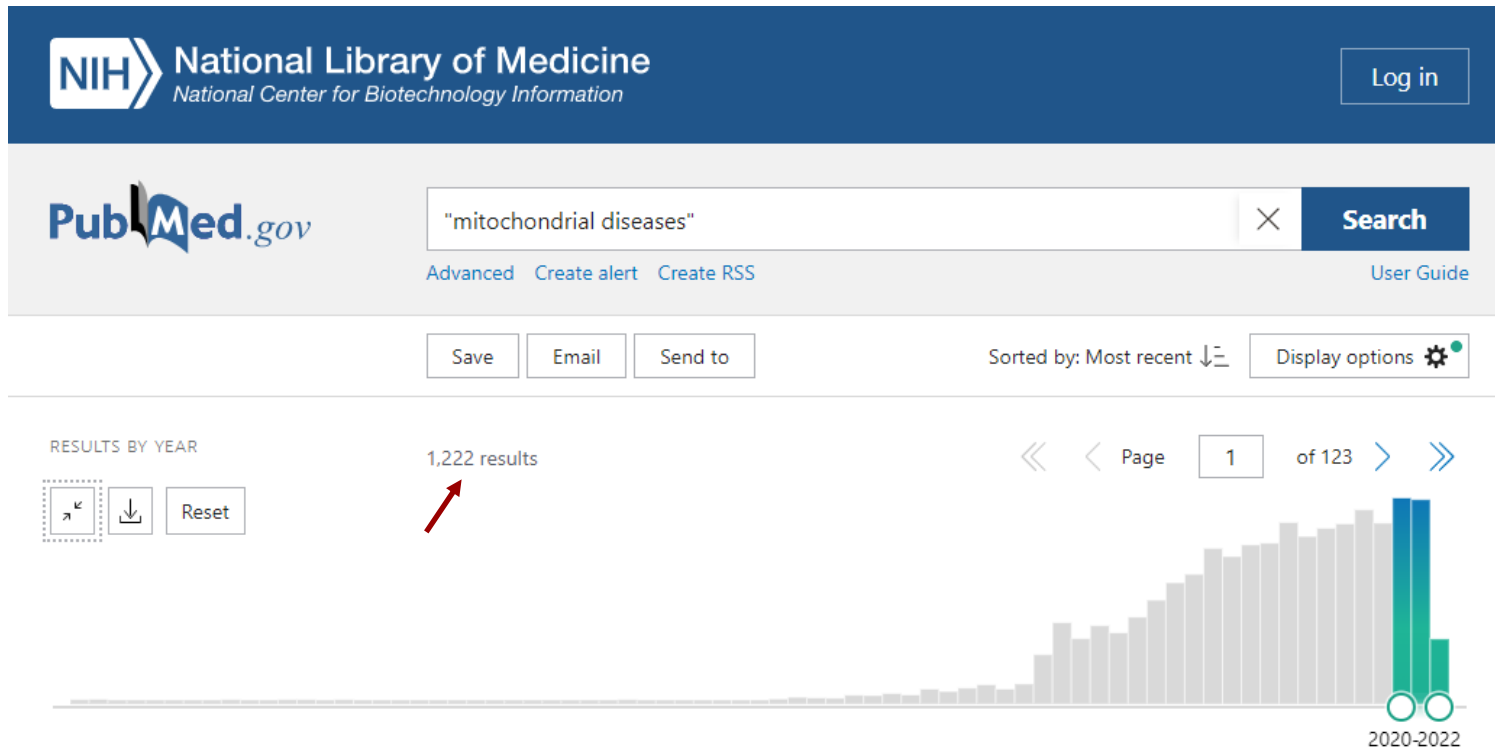
A portrait of a middle-aged man with dark hair, wearing a light-colored trench coat over a striped shirt and a dark tie. He is looking directly at the camera with a slight smile. The background is dark and out of focus.

Rolf Luft, MD, PhD (1914-2007)



Chinnery PF, EMBO Molecular Medicine 2015

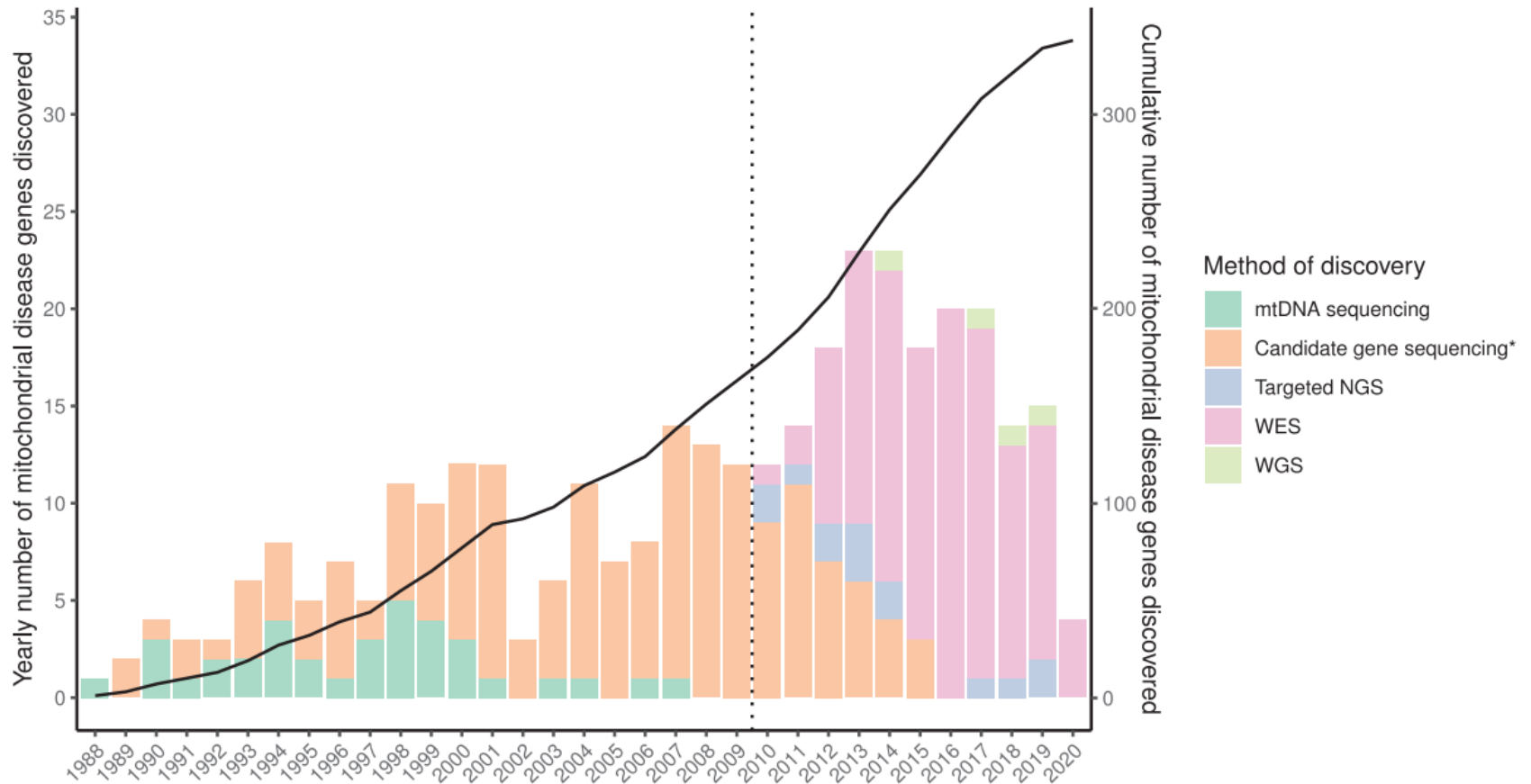
Mitochondrial Diseases



Mitochondrial Diseases



S.L. Stenton and H. Prokisch / EBioMedicine 56 (2020) 102784



Mitochondrial Diseases

Pathogenesis



TISSUE-SPECIFIC STEM CELLS



Mitochondria in neurogenesis: Implications for mitochondrial diseases

Dario Brunetti^{1,2} | Werner Dykstra³ | Stephanie Le⁴ | Annika Zink⁴ | Alessandro Prigione^{3,4}

Nature 524, 234–238 (2015)

Metabolic rescue in pluripotent cells from patients with mtDNA disease

Hong Ma^{1,2}, Clifford D. L. Holmes³, Jun Wu⁴, Robert Morey⁵, Sergio Mora-Castilla⁶, Alejandro Ocampo⁶, Li Ma⁴, Joanna Poulton⁶, Xinjian Wang⁷, Rifat Ahmed⁷, Jianjun Kang⁷, Yeonmi Lee⁷, Tomonari Hayama⁷, Ying Li⁷, Crystal Van Dyken^{1,2}, Nuria Martí Gutierrez^{1,2}, Rebecca Tippler-Hedges^{1,2}, Amy Koski^{1,2}, Nargiz Mitalipov^{1,2}, Paula Amato⁸, Don P. Wolf², Taosheng Huang², Andre Terzic², Louise C. Laurent², Juan Carlos Izpisua Belmonte² & Shoukhrat Mitalipov^{1,2}

New Results

Follow this preprint

Human iPSC-derived cerebral organoids model features of Leigh Syndrome and reveal abnormal corticogenesis

Alejandra I. Romero-Morales, Gabriella L. Robertson, Anuj Rastogi, Megan L. Rasmussen, Hoor Temuri, Gregory Scott McElroy, Ram Prosad Chakrabarty, Lawrence Hsu, Paula M. Almonacid, Bryan A. Millis, Navdeep S. Chandel, Jean-Philippe Cartiailler, Vivian Gama
doi: <https://doi.org/10.1101/2020.04.21.054361>

Cell Death and Disease (2020)11:182

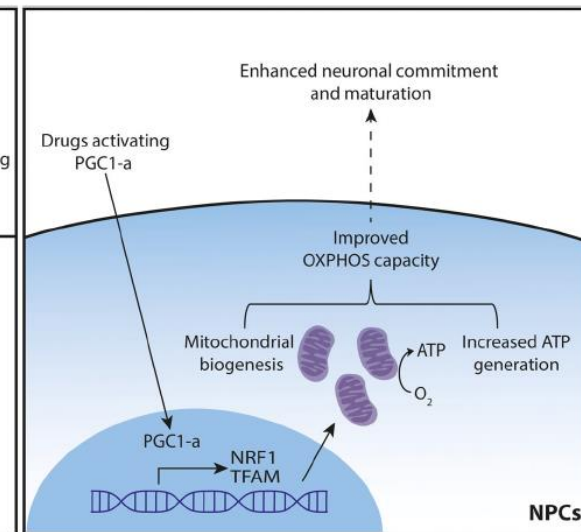
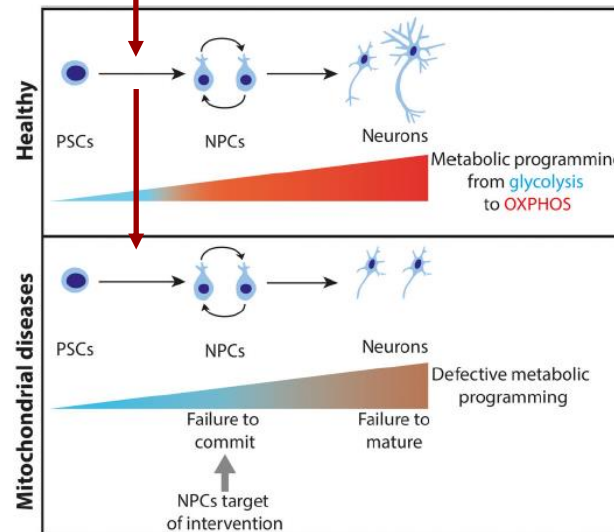
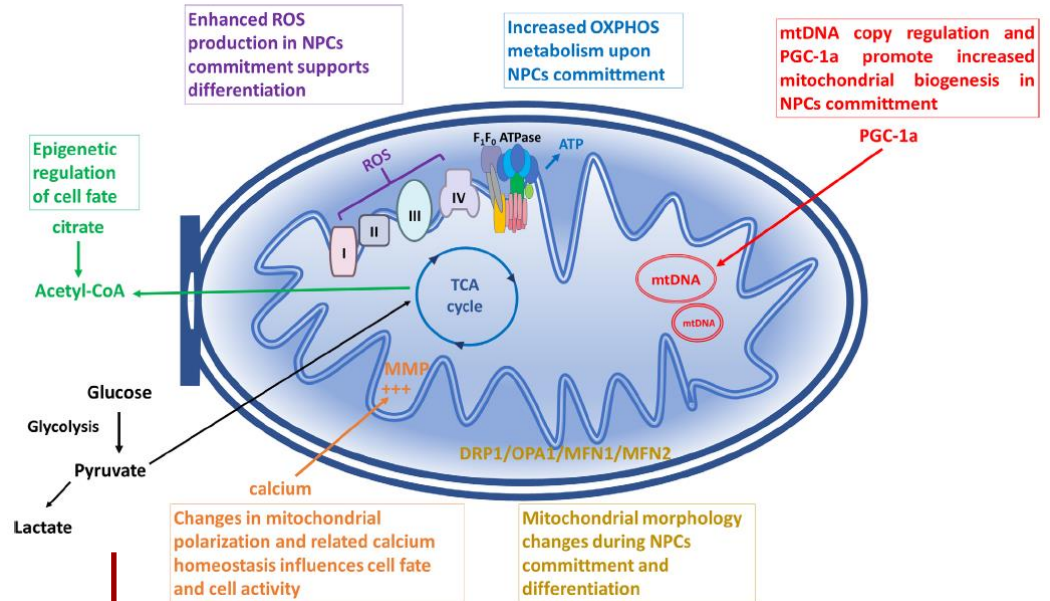
Organoid cultures of MELAS neural cells reveal hyperactive Notch signaling that impacts neurodevelopment

Winanto¹, Zi Jian Khong^{1,2}, Boon-Seng Soh^{1,3,4}, Yong Fan⁴ and Shi-Yan Ng^{1,4,5,6}

Defective metabolic programming impairs early neuronal morphogenesis in neural cultures and an organoid model of Leigh syndrome

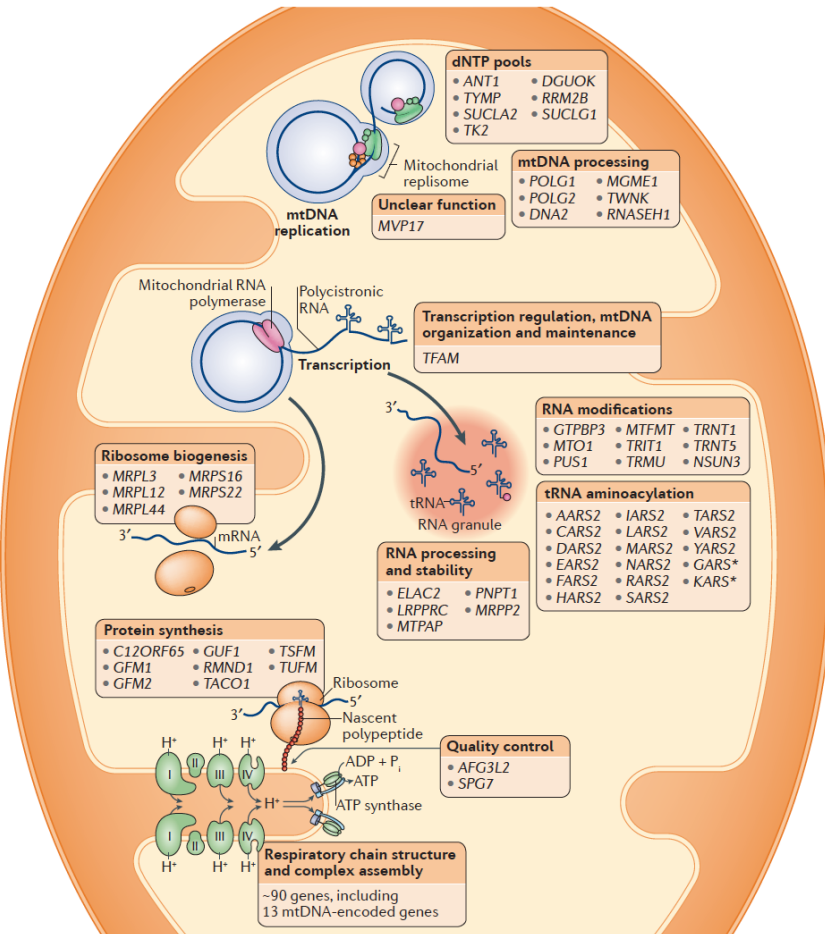
Gizem Inak, Agnieszka Rybak-Wolf, ... Alessandro Prigione

Nature Communications 12, Article number: 1929 (2021) | Cite this article



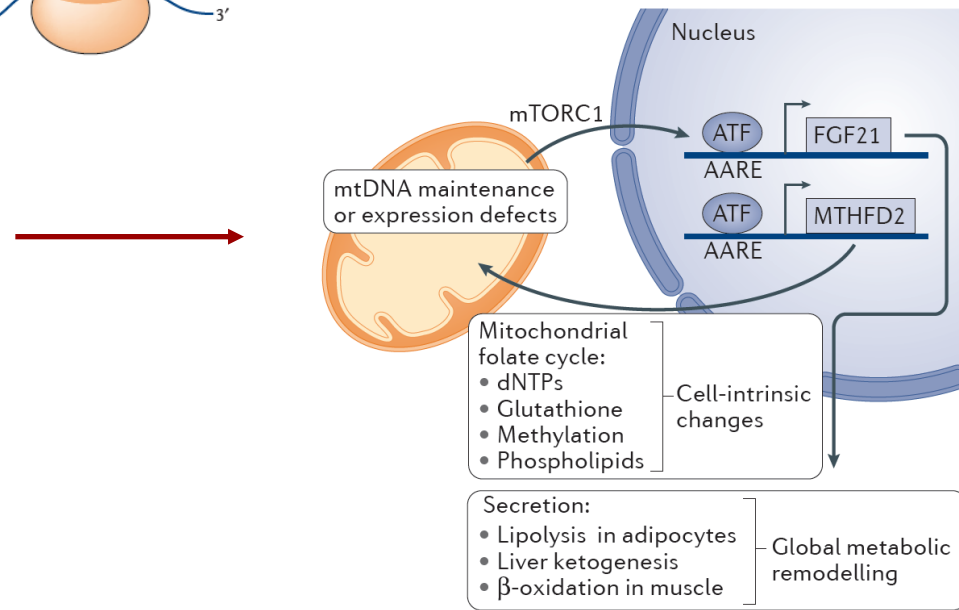
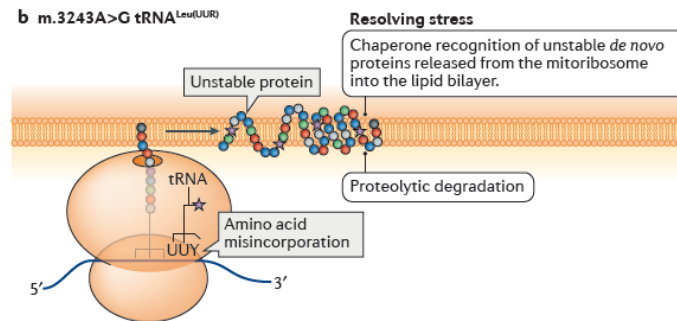
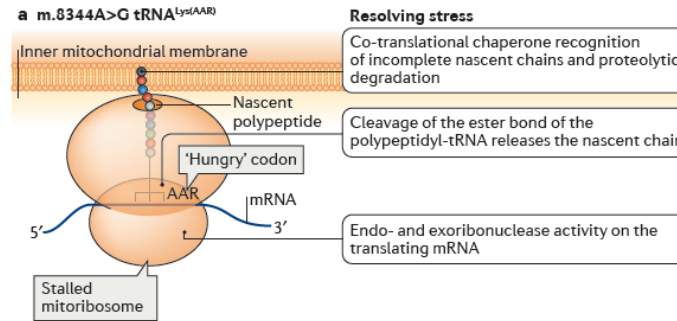
Mitochondrial diseases: the contribution of organelle stress responses to pathology

Anu Suomalainen¹⁻³ and Brendan J. Battersby⁴



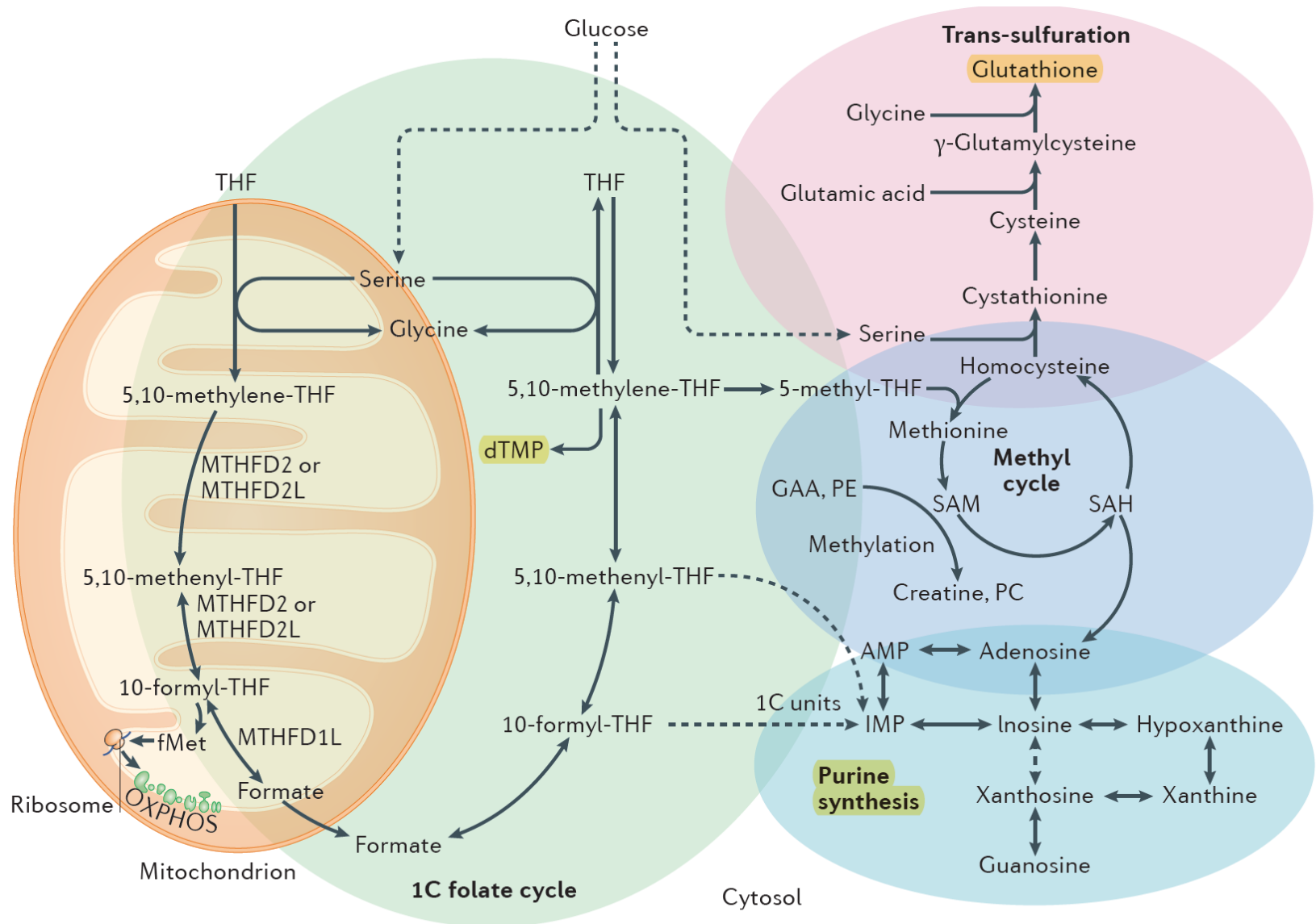
Mitochondrial Diseases

Pathogenesis



Mitochondrial Diseases

Pathogenesis



dTMP, deoxythymidine monophosphate

Mitochondrial Diseases

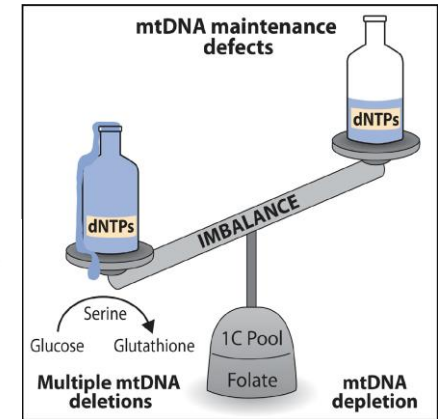
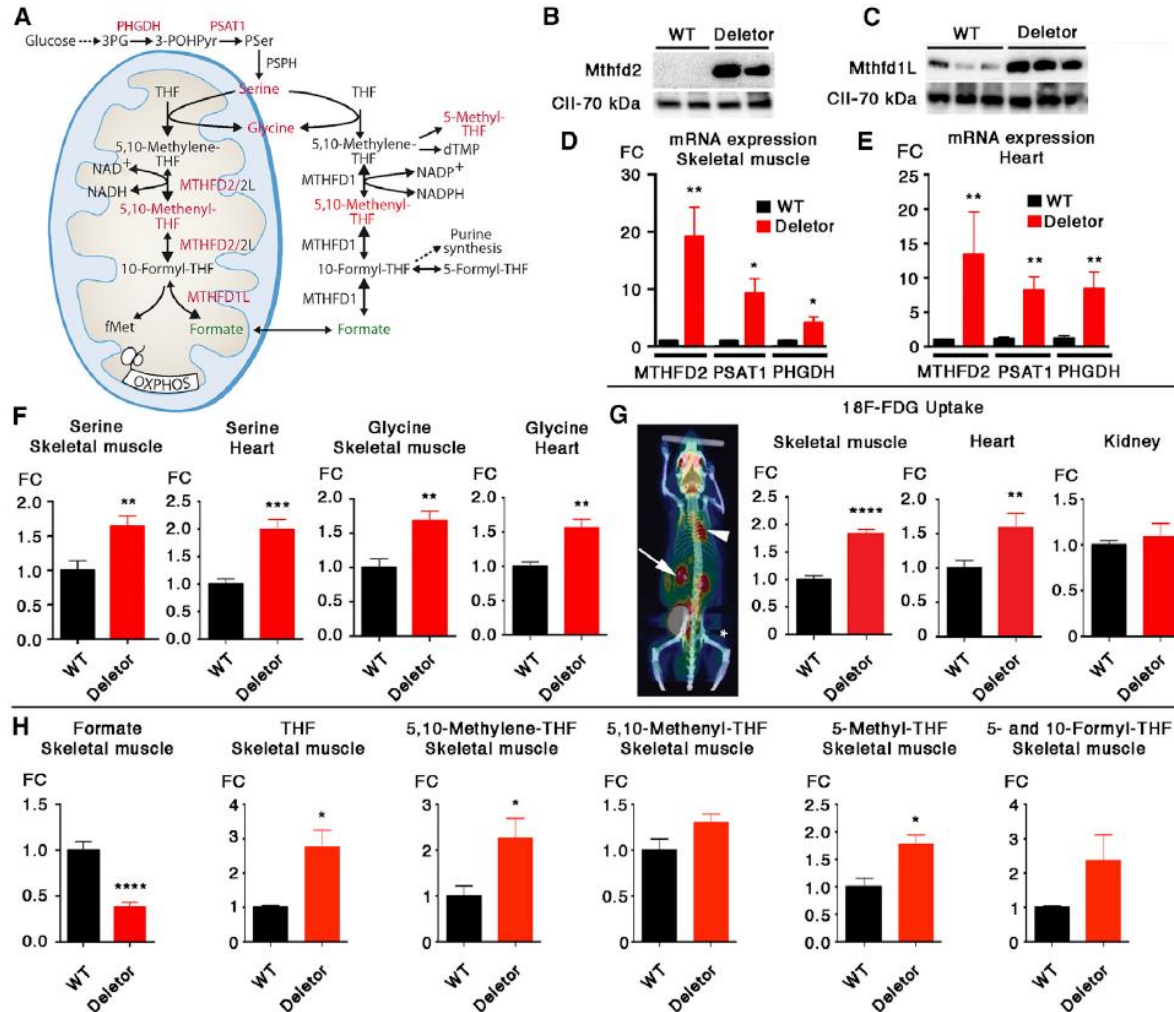
Pathogenesis



Cell Metabolism 23, 635–648, April 12, 2016

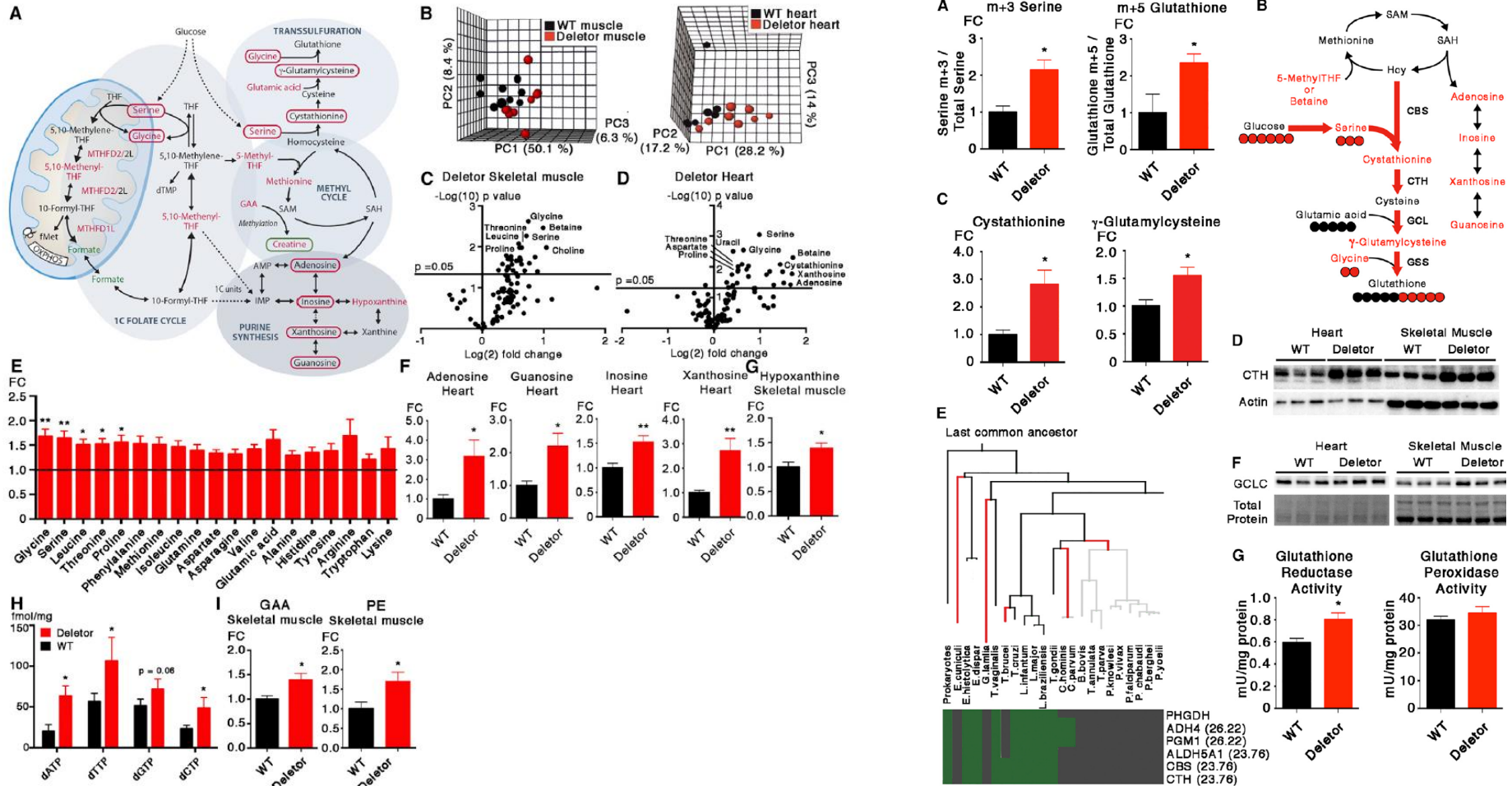
Mitochondrial DNA Replication Defects Disturb Cellular dNTP Pools and Remodel One-Carbon Metabolism

Joni Nikkanen,¹ Saara Forsström,¹ Liliya Euro,¹ Ilse Paetau,¹ Rebecca A. Kohnz,² Liya Wang,³ Dmitri Chilov,¹ Jenni Viinamäki,⁴ Anne Roivainen,^{5,6} Päivi Marjamäki,^{5,6} Heidi Liljenbäck,^{5,6} Sofia Ahola,¹ Jana Buzkova,¹ Mügen Terzioğlu,¹ Nahid A. Khan,¹ Sini Pimes-Karhu,¹ Anders Paetau,⁷ Tuula Lönnqvist,⁸ Antti Sajantila,⁴ Pirjo Isohanni,^{1,8} Henna Tyynismaa,^{1,10} Daniel K. Nomura,² Brendan J. Battersby,¹ Vidya Velagapudi,¹¹ Christopher J. Carroll,^{1,2} and Anu Suomalainen^{1,9,12,*}



Mitochondrial Diseases

Pathogenesis



Mitochondrial Diseases

Pathogenesis



Article

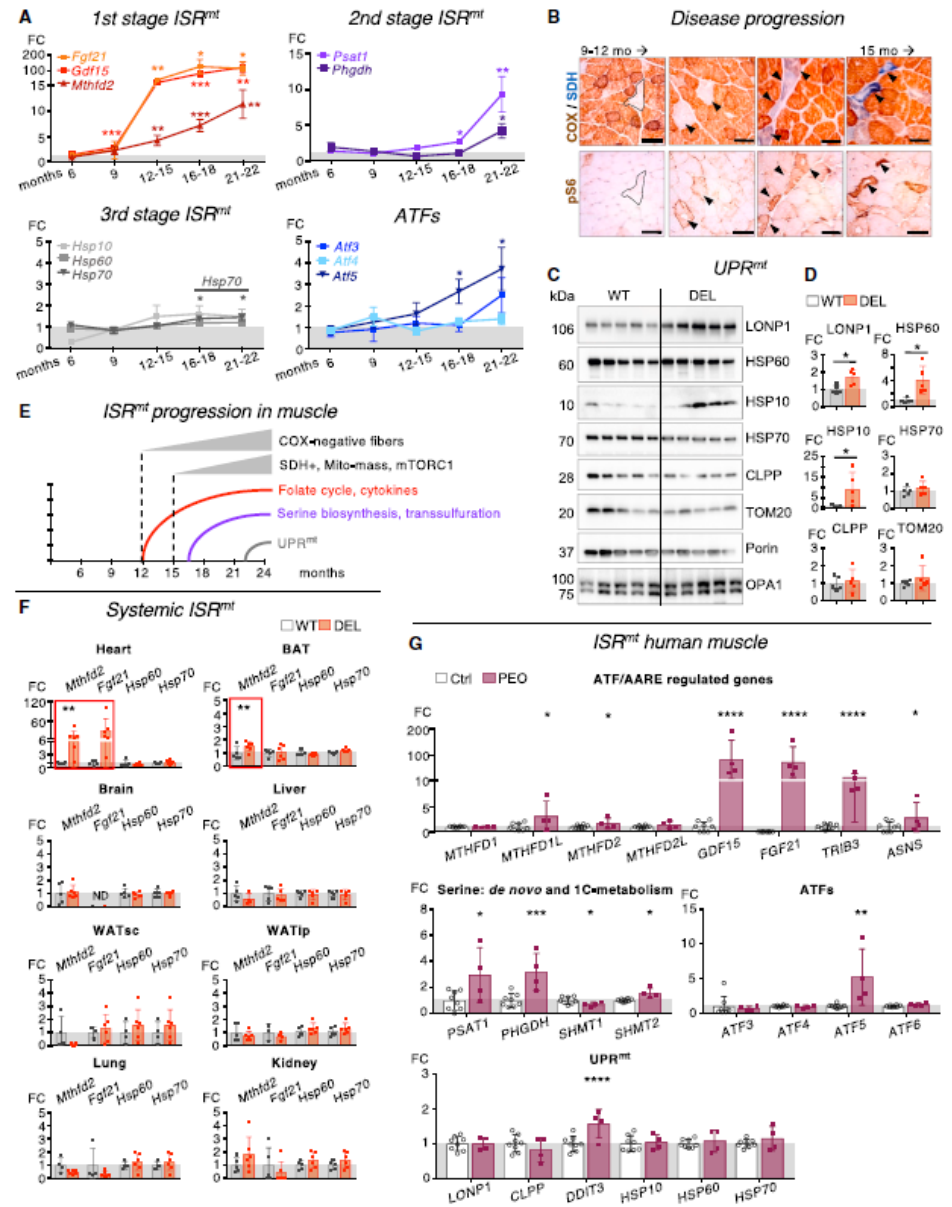
Cell Metabolism

Fibroblast Growth Factor 21 Drives Dynamics of Local and Systemic Stress Responses in Mitochondrial Myopathy with mtDNA Deletions

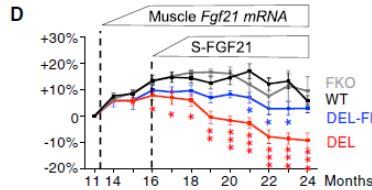
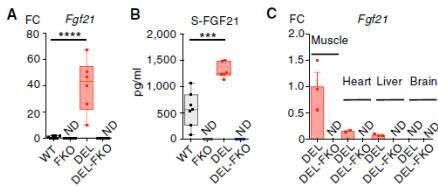
Saara Forsström,^{1,11} Christopher B. Jackson,^{1,11} Christopher J. Carroll,^{1,12} Mervi Kuronen,¹ Eija Pirinen,³ Swagat Pradhan,¹ Anastasia Marmyleva,¹ Mari Auranen,⁴ Iida-Marja Kleine,¹ Nahid A. Khan,¹ Anne Roivainen,^{5,6} Paivi Marjamäki,¹ Heidi Liljenbäck,^{1,2} Liya Wang,⁷ Brendan J. Battersby,⁸ Uwe Richter,⁸ Vidya Velagapudi,⁹ Joni Nikkanen,¹ Liliya Euro,¹ and Anu Suomalainen^{1,4,10,12,*}

Mitochondrial stress responses are

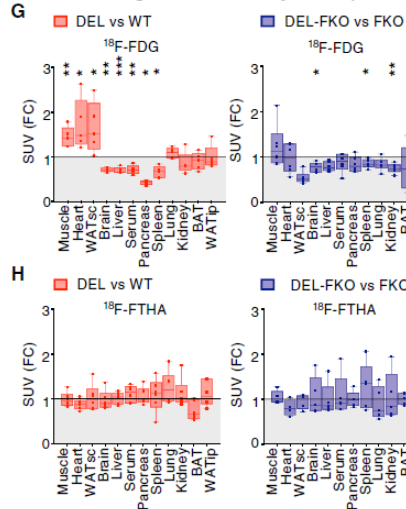
- **only partially conserved in species**
- **not generalizable** between organisms or even mammalian cell types
- **FGF21** is a key mediator of metabolic remodeling and progression of ISRmt locally and systemically



FGF21 expression in mouse lines



In vivo glucose and fatty acid uptake



Mitochondrial Diseases

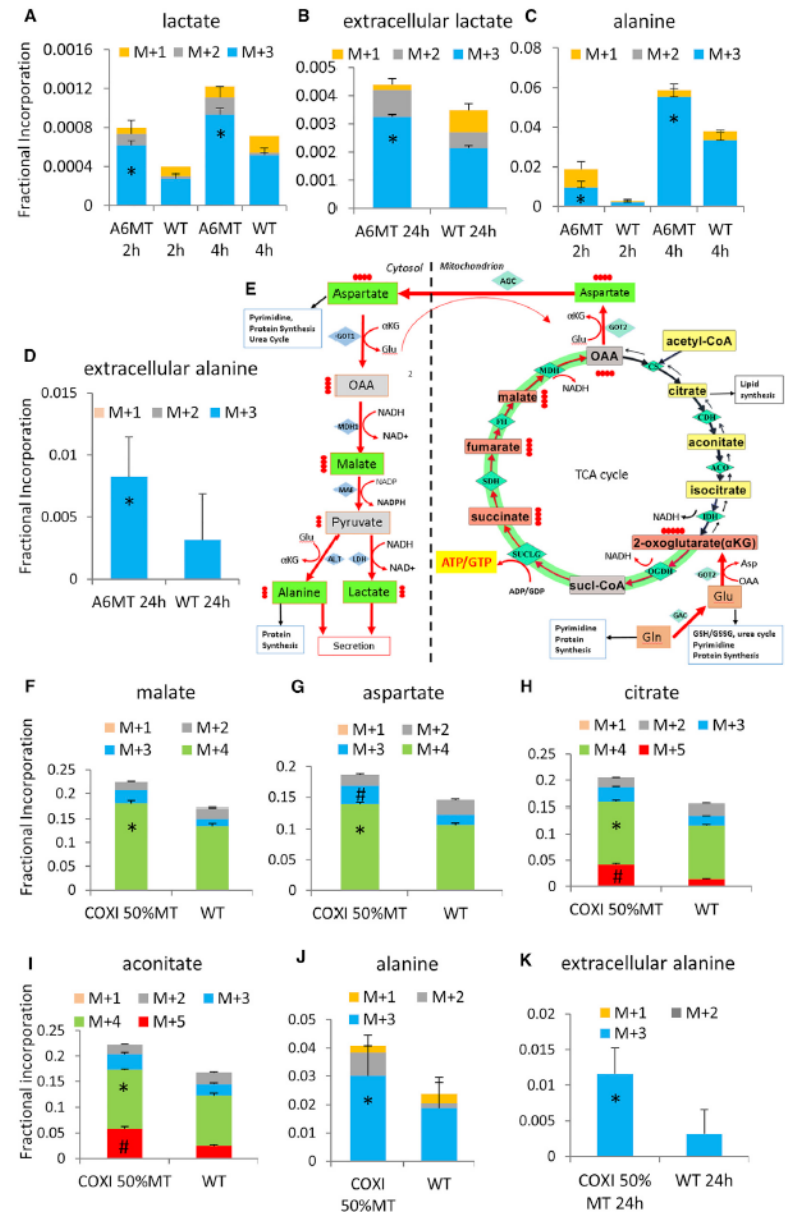
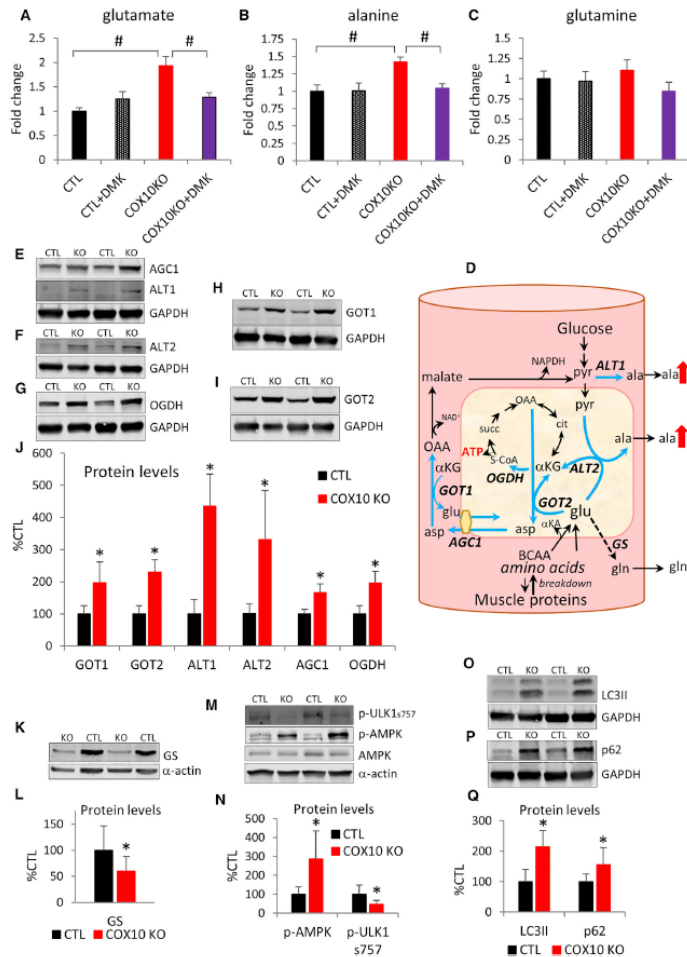
Pathogenesis

Article

Cell Metabolism

Rewiring of Glutamine Metabolism Is a Bioenergetic Adaptation of Human Cells with Mitochondrial DNA Mutations

Qiuying Chen,^{2,8} Kathyne Kirk,^{1,8} Yevgeniya I. Shurubor,¹ Dazhi Zhao,¹ Andrea J. Arreguin,¹ Ifrah Shahi,¹ Federica Valsecchi,¹ Guido Primiano,³ Elizabeth L. Calder,⁴ Valerio Carelli,^{5,6} Travis T. Denton,⁷ M. Flint Beal,¹ Steven S. Gross,² Giovanni Manfredi,^{1,5} and Mariena D'Aurelio^{1,8,*}





Lancet Neurol 2011; 10: 806–18

FGF-21 as a biomarker for muscle-manifesting mitochondrial respiratory chain deficiencies: a diagnostic study

ANN NEUROL 2015;78:814–823

Growth Differentiation Factor 15 as a Useful Biomarker for Mitochondrial Disorders



RESEARCH ARTICLE

Accuracy of FGF-21 and GDF-15 for the diagnosis of mitochondrial disorders: A meta-analysis

Yan Lin¹, Kunqian Ji¹, Xiaotian Ma², Shuangwu Liu¹, Wei Li¹, Yuying Zhao¹ & Chuanzhu Yan^{1,2,3} 











Biomarkers

















Estimates (95% CI)

	FGF-21	GDF-15
Number of included studies	5	7
Number of subjects	718	845
Sensitivity	0.71 (0.53, 0.84)	0.83 (0.65, 0.92)
Specificity	0.88 (0.82, 0.93)	0.92 (0.84, 0.96)
Positive likelihood ratio	6.10 (3.40, 10.70)	9.90 (4.60, 21.20)
Negative likelihood ratio	0.33 (0.19, 0.58)	0.19 (0.08, 0.42)
Diagnostic odds ratio	18.00 (6.00, 54.00)	52.00 (13.00, 205.00)
AUC	0.90 (0.87, 0.92)	0.94 (0.92, 0.96)

A. FGF-21

Study	TP	FP	FN	TN	Sensitivity (95% CI)	Specificity (95% CI)	Sensitivity (95% CI)	Specificity (95% CI)
Davis RL (2016)	37	6	17	60	0.69 [0.54, 0.80]	0.91 [0.81, 0.97]		
Montero R (2016)	25	1	23	32	0.52 [0.37, 0.67]	0.97 [0.84, 1.00]		
Suomalainen A (2011)	62	6	5	68	0.93 [0.83, 0.98]	0.92 [0.83, 0.97]		
Tsygankova PG (2019)	62	14	60	46	0.51 [0.42, 0.60]	0.77 [0.64, 0.87]		
Yatsuga S (2015)	37	18	11	128	0.77 [0.63, 0.88]	0.88 [0.81, 0.93]		

B. GDF-15

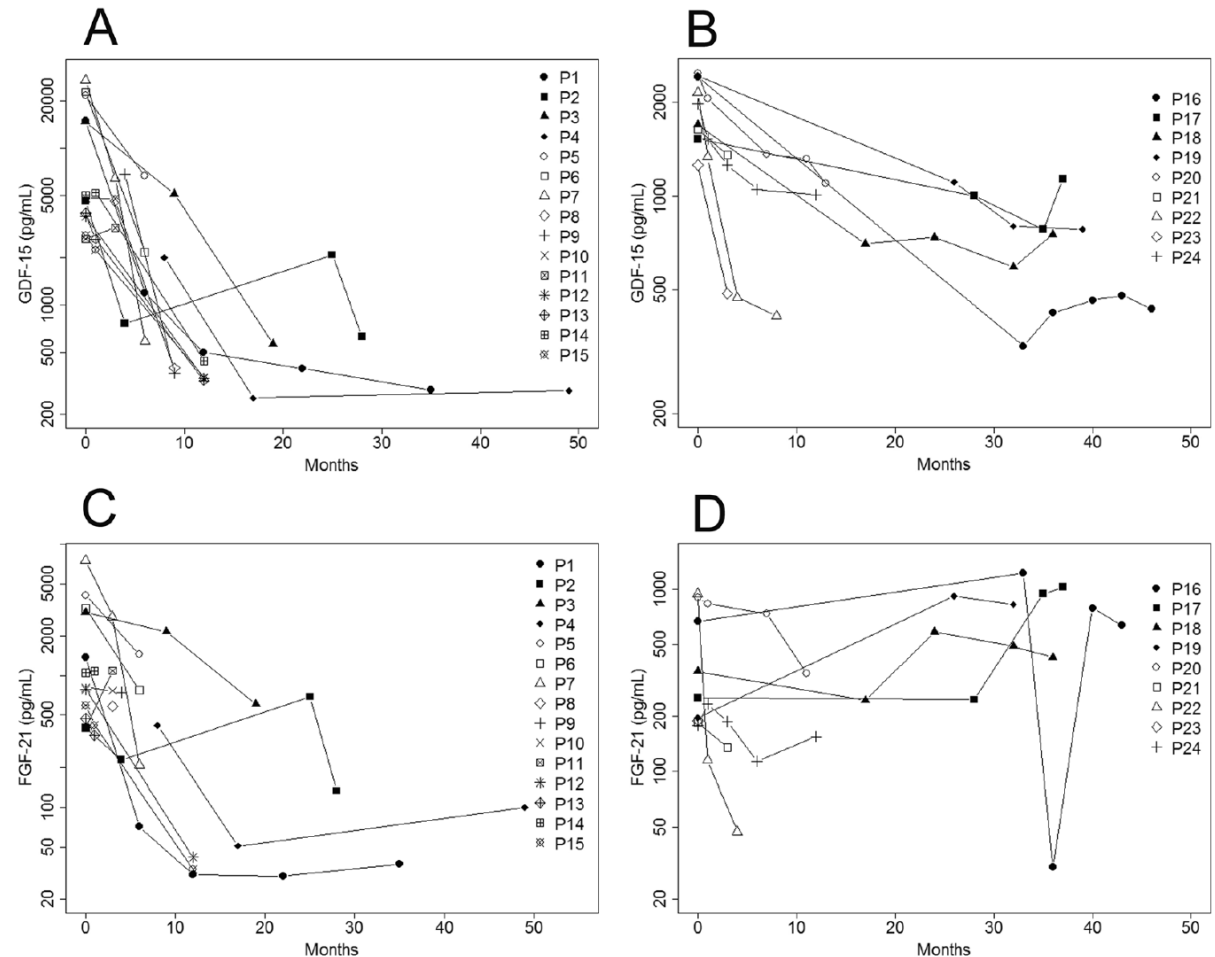
Study	TP	FP	FN	TN	Sensitivity (95% CI)	Specificity (95% CI)	Sensitivity (95% CI)	Specificity (95% CI)
Davis RL (2016)	42	4	12	62	0.78 [0.64, 0.88]	0.94 [0.85, 0.98]		
Ji X (2016)	41	4	1	46	0.98 [0.87, 1.00]	0.92 [0.81, 0.98]		
Koene S (2015)	51	1	46	29	0.53 [0.42, 0.63]	0.97 [0.83, 1.00]		
Montero R (2016)	33	3	15	30	0.69 [0.54, 0.81]	0.91 [0.76, 0.98]		
Poulsen NS (2019)	23	1	5	20	0.82 [0.63, 0.94]	0.95 [0.76, 1.00]		
Tsygankova PG (2019)	81	22	41	38	0.66 [0.57, 0.75]	0.63 [0.50, 0.75]		
Yatsuga S (2015)	47	7	1	139	0.98 [0.89, 1.00]	0.95 [0.90, 0.98]		



Growth Differentiation Factor 15 is a potential biomarker of therapeutic response for TK2 deficient myopathy


Cristina Dominguez-Gonzalez^{1,2,3}, Carmen Badosa⁴, Marcos Madruga-Garrido⁵, Itxaso Marti⁶, Carmen Paradas^{7,8}, Carlos Orteza⁴, Jordi Diaz-Manera^{3,9}, Andres Berardo¹¹, Jorge Alonso-Pérez⁹, Selena Trifunov⁴, Daniel Cuadras¹¹, Susana G. Kalko¹², Cora Blázquez Bermejo^{3,13}, Yolanda Cámara^{3,13}, Ramon Martí^{3,13}, Fabiola Mavillard^{7,8}, Miguel A. Martí Julio Montoya^{3,14}, Eduardo Ruiz-Pesini^{3,14}, Joan Villarroya^{15,16}, Raquel Montero^{3,17}, Francesc Villarroya^{15,16}, Rafael Artuch^{3,17}, Michio Hirano¹⁰, Andrés Nascimento^{3,4} & Cecilia Jimenez-Mallebrera^{3,4,18} 

- **24 patients** with TK2 deficiency
- Significant correlation between basal GDF-15 and **6MWT**
- During treatments with oral deoxynucleosides, **GDF15 significantly declined**





Diagnostic value of serum biomarkers FGF21 and GDF15 compared to muscle sample in mitochondrial disease

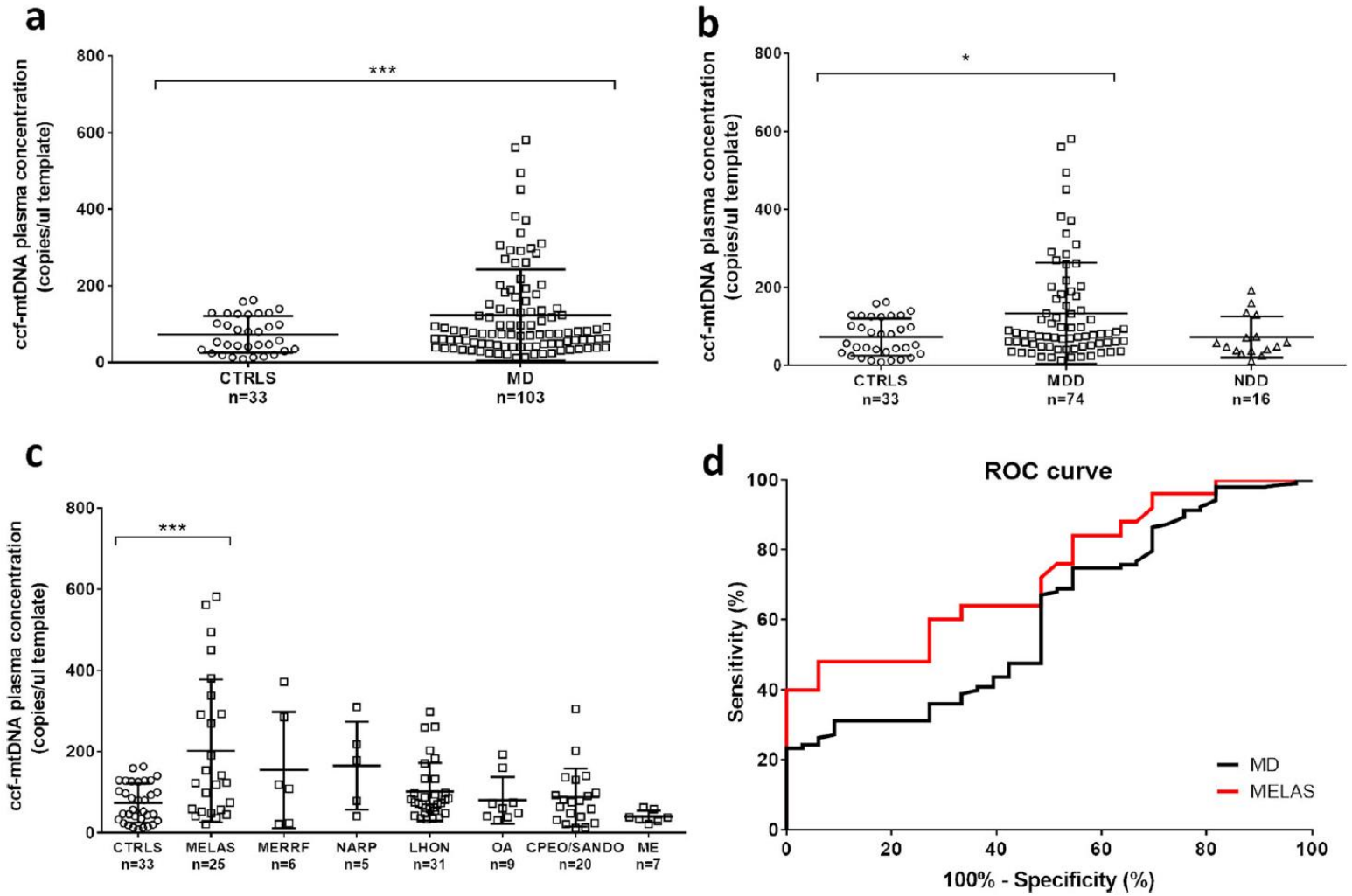
Jenni M. Lehtonen¹ | Mari Auranen^{1,2} | Niklas Darin³ | Kalliopi Sofou³ |
Laurence Bindoff^{4,5} | Omar Hikmat^{4,6} | Johanna Uusimaa⁷ | Päivi Vieira⁷ |
Már Tulinius³ | Tuula Lönnqvist⁸ | Irenaeus F. de Co^{9,10} |
Anu Suomalainen^{1,11} | Pirjo Isohanni^{1,8} 

- **194 suspected mitochondrial disorders** (88 children and 106 adults)
- **FGF21 and GDF15 identified 62%** of patients with genetically verified MD (**82% muscle manifesting**)
- Serum biomarkers pointed to mitochondrial disease in **69% patients who had no diagnostic findings in the muscle sample**
- **FGF21 and GDF15** was highly restricted to **muscle-manifesting mitochondrial diseases** caused by mitochondrial translation defect or mtDNA deletions
- FGF21 and GDF15 together as **first-line diagnostic tools in patients with muscle involvement**; they can be used in all patients with a suspicion of mitochondrial disease, although in pure encephalopathies biomarkers often remain normal
- Analysis of serum biomarkers complement but **do not entirely remove the need for muscle biopsy**
- Biomarkers might be useful in **evaluation of genetic findings**, for example, variants of unknown significance



Expanding and validating the biomarkers for mitochondrial diseases

Alessandra Maresca¹ · Valentina Del Dotto² · Martina Romagnoli¹ · Chiara La Morgia^{1,2} · Lidia Di Vito¹ · Marianonietta Capristo¹ · Maria Lucia Valentino^{1,2} · Valerio Carelli^{1,2}  · the ER-MITO Study Group



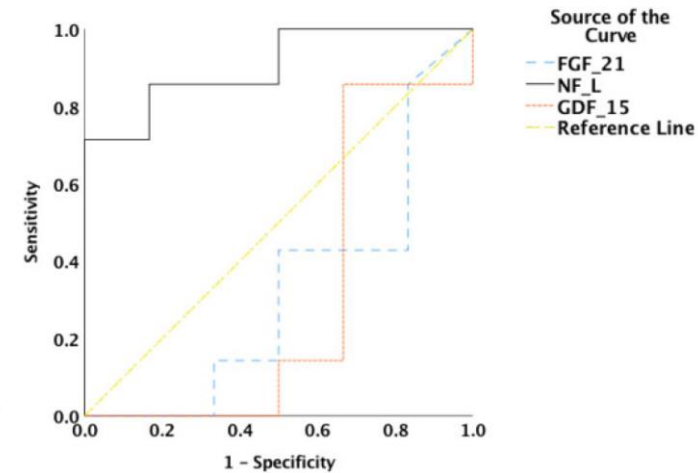
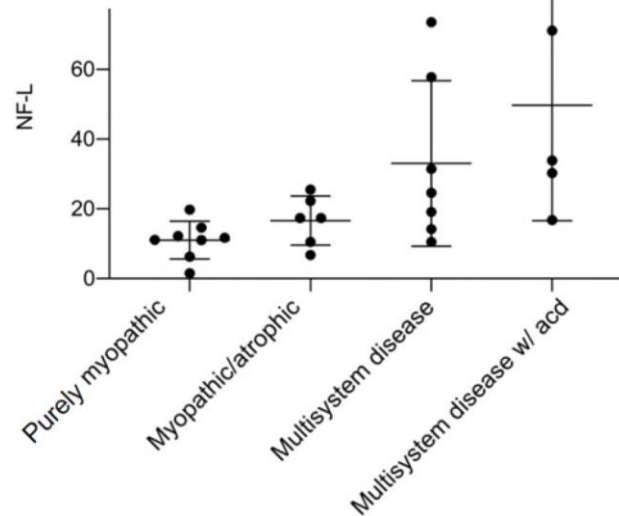
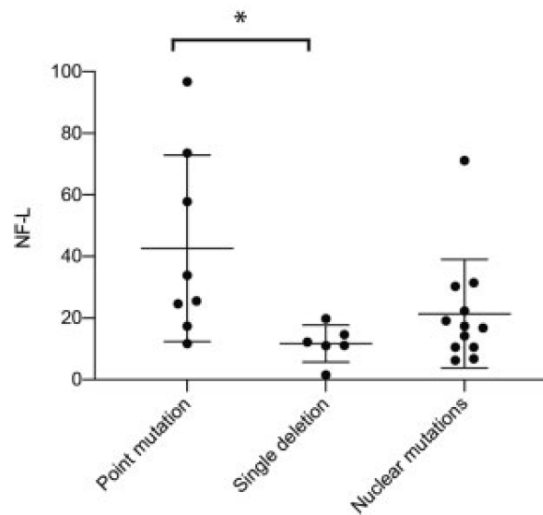


BRAIN COMMUNICATIONS 2021:

Serum biomarkers in primary mitochondrial disorders

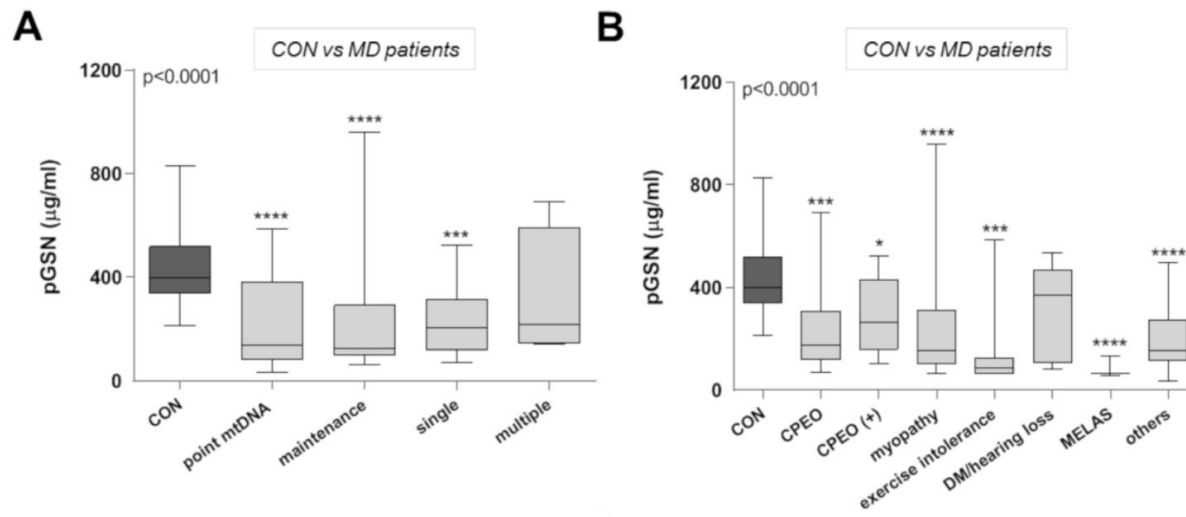
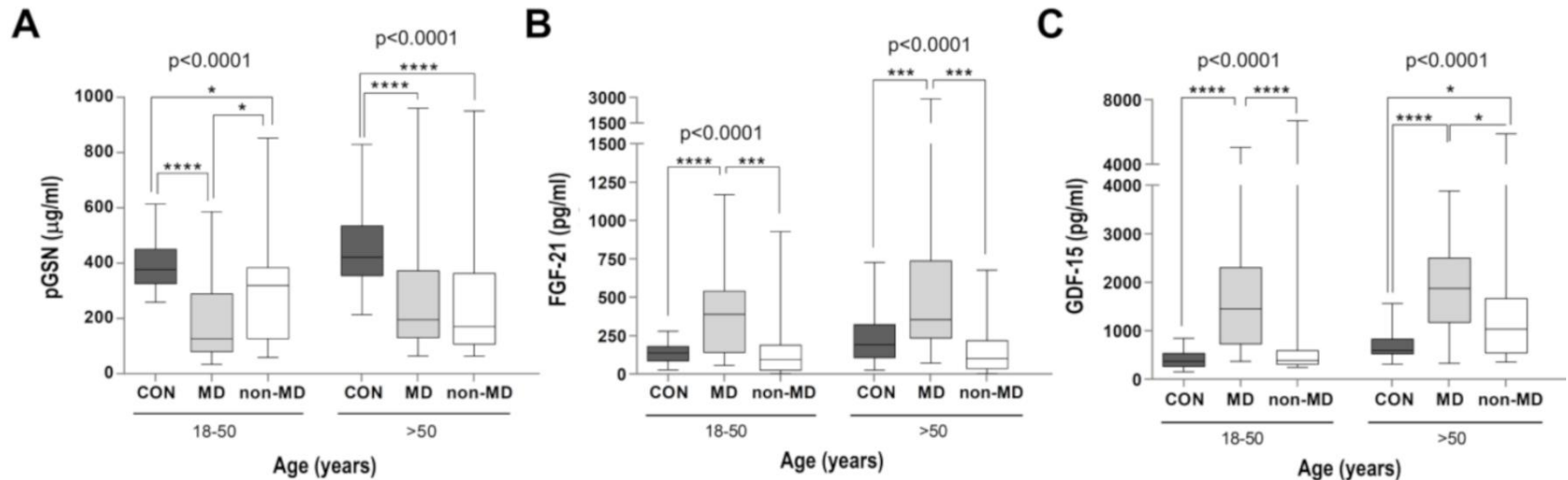
Kristin N. Varhaug,^{1,2} Omar Hikmat,^{2,3} Hanne Linda Nakkestad,^{1,4} Christian A. Vedeler^{1,2,4}
and Laurence A. Bindoff^{1,2,4}

	MtDNA point mutations	N	Single deletions	Nuclear gene mutations	N
Genetic diagnose	8344 A>G	2		<i>POLG</i>	7
	3243 A>G	4		<i>TWINKL</i>	2
	13271 T>C	1		<i>PITRM1</i>	1
	5556 G>C	1		<i>DHX30</i>	1
				<i>ICSU</i>	1
Age (mean years)	58		43	47	
Gender (female %)	63 %		100 %	83 %	
Total		8	6		12



Plasma Gelsolin Reinforces the Diagnostic Value of FGF-21 and GDF-15 for Mitochondrial Disorders

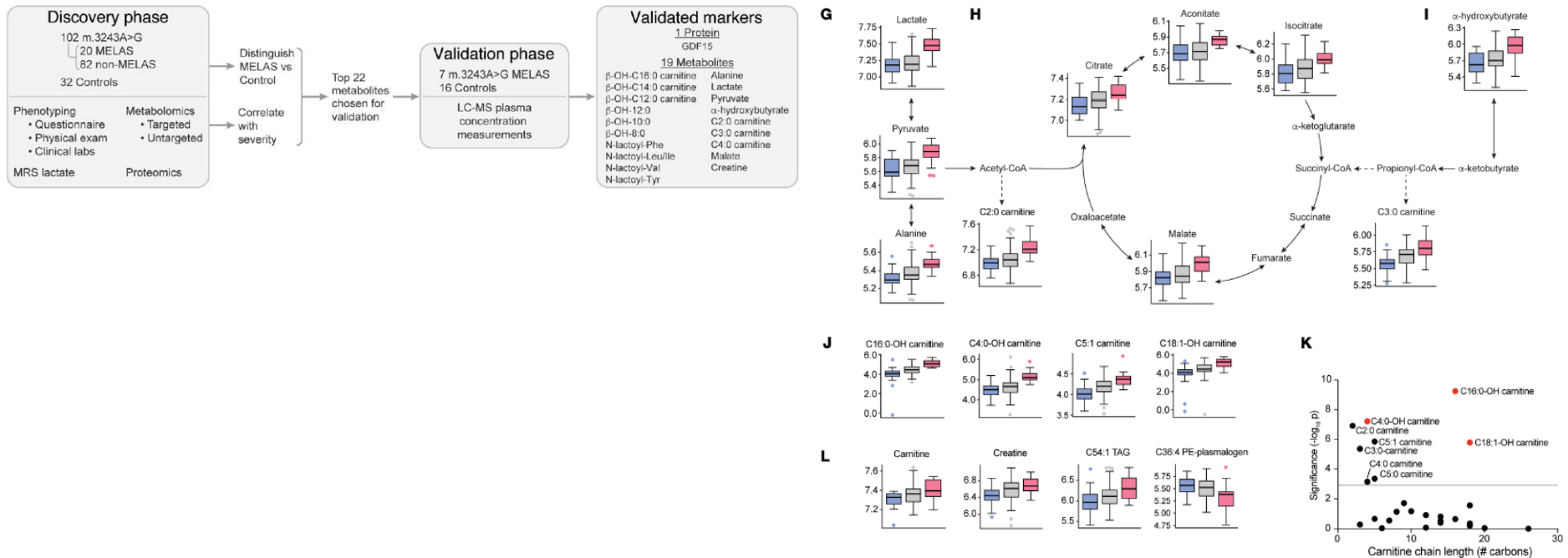
Ana Peñas ¹, Miguel Fernández-De la Torre ¹, Sara Laine-Menéndez ¹, David Lora ^{2,3,4}, María Illescas ¹, Alberto García-Bartolomé ¹, Montserrat Morales-Conejo ^{1,5,6}, Joaquín Arenas ^{1,6}, Miguel A. Martín ^{1,6,7}, María Morán ^{1,6}, Cristina Domínguez-González ^{1,6,8,*} and Cristina Ugalde ^{1,6,*}





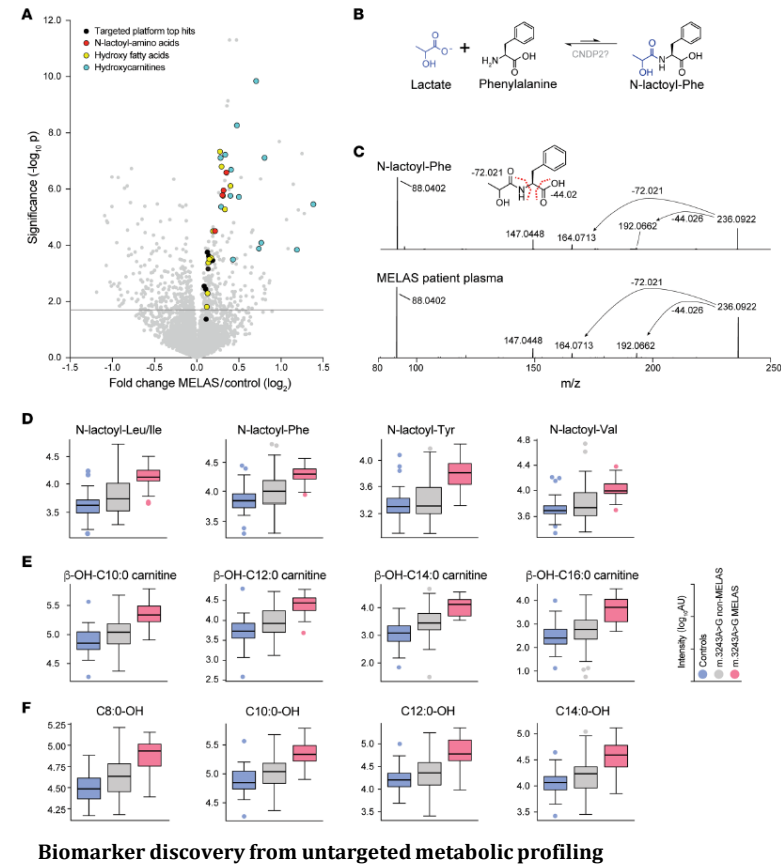
Circulating markers of NADH-reductive stress correlate with mitochondrial disease severity

Rohit Sharma,^{1,2,3,4} Bryn Reinstadler,^{1,2,3,4} Kristin Engelstad,⁵ Owen S. Skinner,^{1,2,3,4} Erin Stackowitz,⁵ Ronald G. Haller,^{6,7} Clary B. Clish,⁴ Kerry Pierce,⁴ Melissa A. Walker,^{1,2,3,4,8} Robert Fryer,⁹ Devin Oglesbee,⁹ Xiangling Mao,¹⁰ Dikoma C. Shungu,¹⁰ Ashok Khatri,¹¹ Michio Hirano,⁵ Darryl C. De Vivo,⁵ and Vamsi K. Mootha^{1,2,3,4}

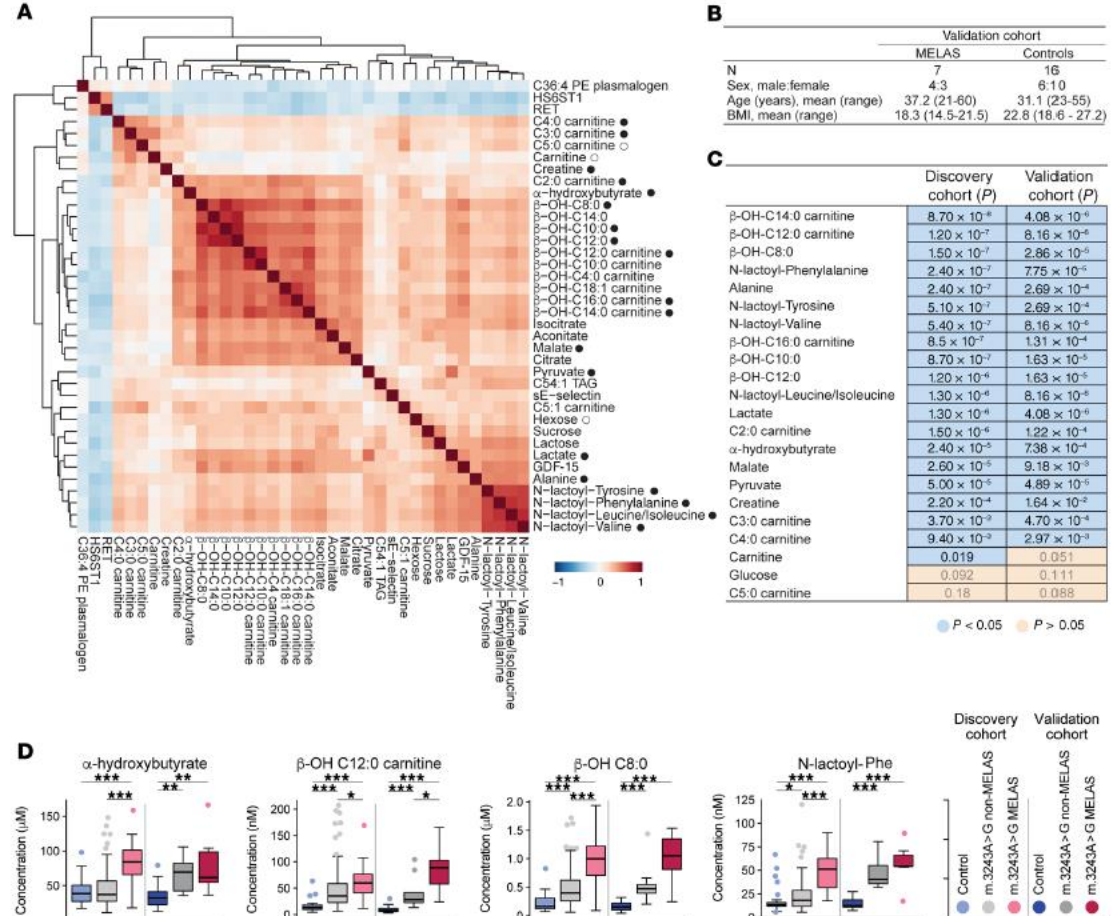


Comparison of 1310 proteins and 376 targeted metabolites in plasma of patients with MELAS and controls.

Biomarkers

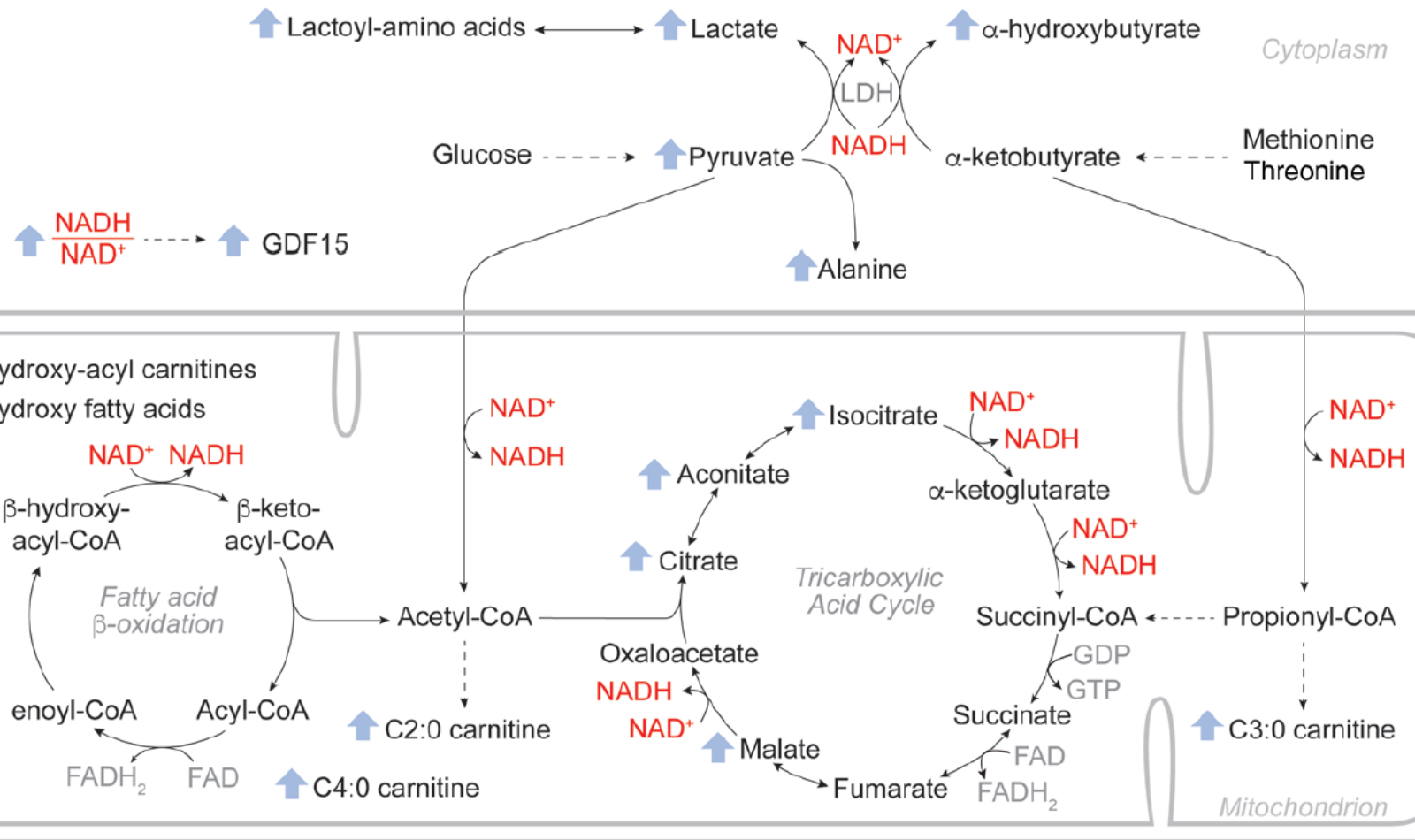


Biomarker discovery from untargeted metabolic profiling



Correlations among most discriminating analytes and validation of top metabolites

Biomarkers



NADH-reductive stress drives the metabolic signature in MELAS.

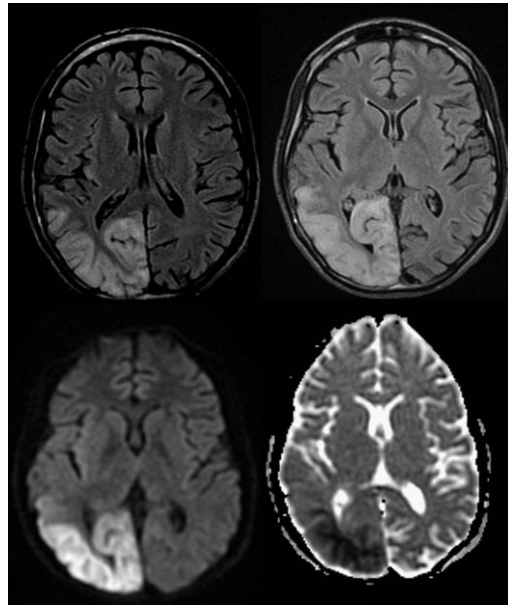
Classical mitochondrial syndromes



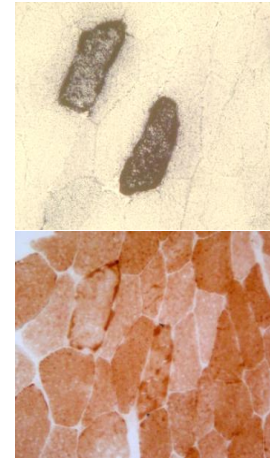
PEO spectrum/Kearns-Sayre syndrome



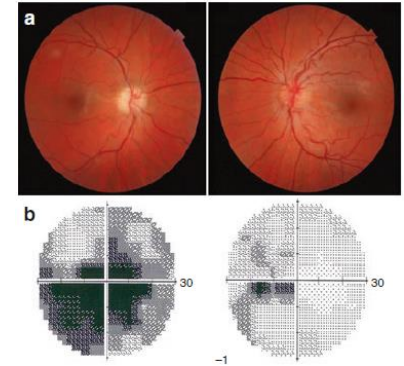
MELAS



MIDD

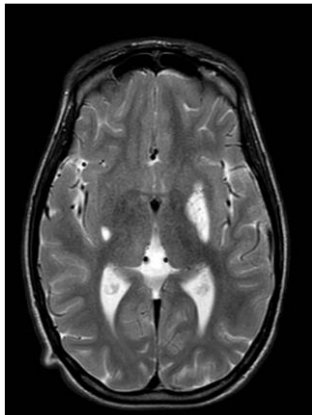


LHON



Carelli V, et al. 2019

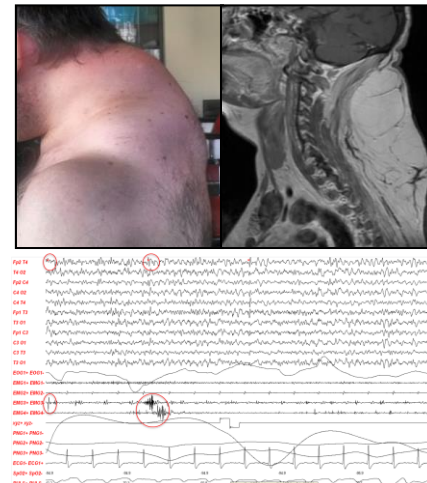
Leigh Syndrome



MNGIE



MERRF



Classical mitochondrial syndromes

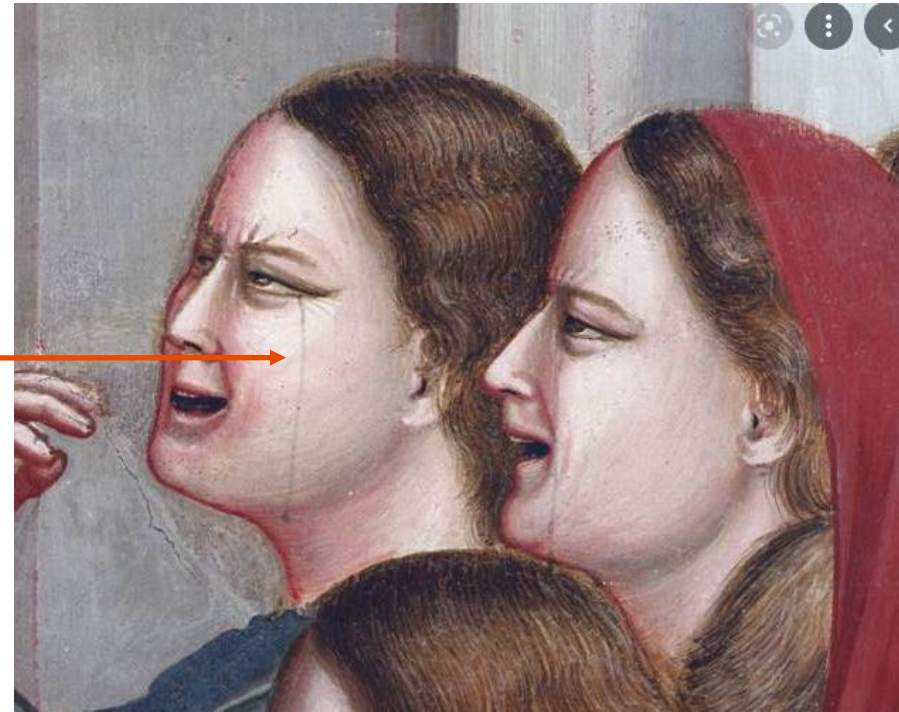


Laocoonte and His Sons, Vatican Museums

"The first time I was in Rome when I was very young, the Pope [Julius II] was told about **the discovery of some very beautiful statues in a vineyard near Santa Maria Maggiore** [on the Esquiline Hill]. The pope ordered one of his officers to run and tell **Giuliano da Sangallo** to go and see them... Since **Michelangelo Buonarroti** was always to be found at our house, my father having summoned him and having assigned him the commission of the Pope's tomb, my father wanted him to come along too... I had climbed down to where the statues were, when immediately **my father said, 'That is the Laocoon, which Pliny mentions.'** Then they dug the hole wider so that that they could pull the statue out. As soon as it was visible everyone started to draw, all the while discoursing on ancient things, chatting about the ones [ancient statues owned by the Medici] in Florence."

Letter of Francesco da Sangallo, quoted in Leonard Barkan, *Unearthing the Past: Archaeology and Aesthetics in the Making of Renaissance Culture* (1999)

Refined phenotypes



The Massacre of the Innocents, Scrovegni Chapel, Padova, Italy

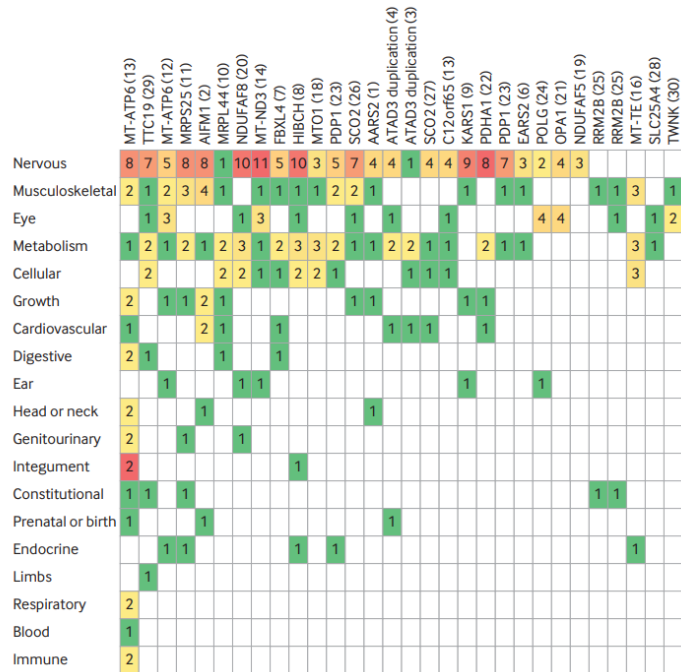
Refined phenotypes



thebmj | BMJ 2021;375:e066288 | doi: 10.1136/bmj-2021-066288

Use of whole genome sequencing to determine genetic basis of suspected mitochondrial disorders: cohort study

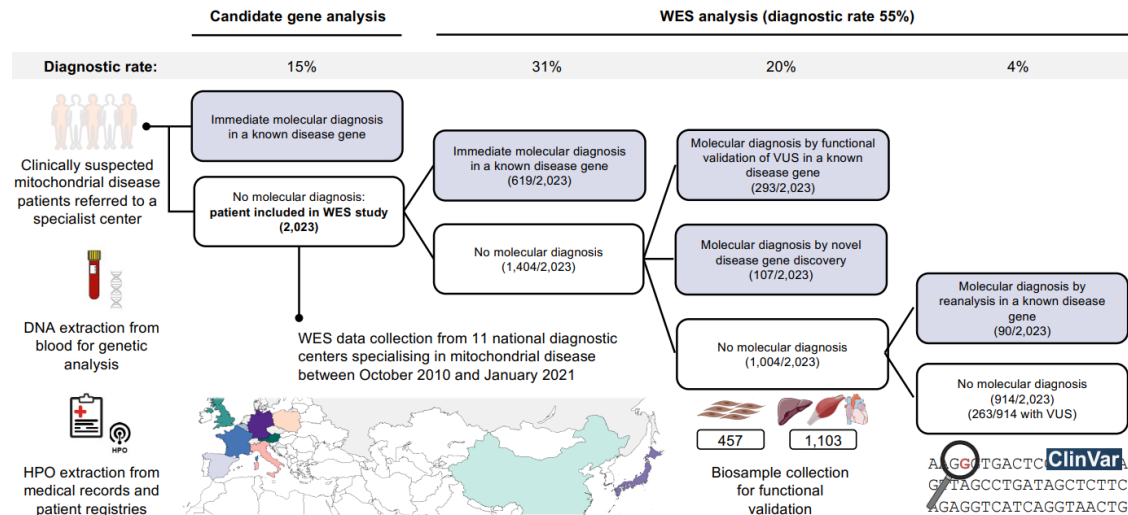
Human Phenotype Ontology (HPO) terms in patients with mitochondrial diagnoses.



Diagnosing pediatric mitochondrial disease: lessons from 2,000 exomes

Sarah L. Stenton, Masaru Shimura, Dorota Piekutowska-Abramczuk, Peter Freisinger, Felix Distelmaier, Johannes A. Mayr, Christine Makowski, Borianna Büchner, Bader Alhaddad, Charlotte L. Alston, Anna Ardisson, Rui Ban, Ivo Barić, Riccardo Berutti, Theresa Brunet, Elzbieta Ciara, Dasha Deen, Julien Gagneur, Daniele Ghezzi, Mirjana Gusic, Tobias B. Haack, Maja Hempel, Ralf A. Husain, Daniela Karall, Stefan Kölker, Urania Kotzaeridou, Thomas Klopstock, Robert Kopajtich, Vassiliki Konstantopoulou, Steffen Liez, Dominic Lenz, Albert Z. Lim, Hanna Mandel, Robert McFarland, Wolfgang Müller-Felber, Gerard Muñoz-Pujol, Akira Ohtake, Yasushi Okazaki, Rikke Olsen, Ewa Pronicka, Angela Pyle, Antonia Ribes, Dariusz Rokicki, René Santer, Manuel Schiff, Markus Schuelke, Dmitrii Smirnov, Wolfgang Sperl, Tim Strom, Frederic Tort, Polina Tsygankova, Rudy van Coster, Patrick Verloo, Jürgen-Christoph von Kleist-Retzow, Ekkehard Wilichowski, Tekla Wolstein, Manting Xu, Vicente Yépez, Michael Zech, Saskia Wortmann, Matias Wagner, Costanza Lamperti, Robert W. Taylor, Fang Fang, Agnès Rötig, Kei Murayama, Thomas Meitinger, Holger Prokisch

doi: <https://doi.org/10.1101/2021.06.21.21259171>



Refined phenotypes



MitoPhen database: a human phenotype ontology-based approach to identify mitochondrial DNA diseases

Thiloka E. Ratnaike^{1,2,3,†}, Daniel Greene^{4,5,†}, Wei Wei^{1,2}, Alba Sanchis-Juan⁴, Katherine R. Schon^{1,2,6}, Jelle van den Amele^{1,2}, Lucy Raymond⁶, Rita Horvath^{6,1,2}, Ernest Turro^{7,†} and Patrick F. Chinnery^{6,1,2,†}

- 89 mtDNA variants, spanning 27 genes
- 676/1352 publications
- Data from 6688 individuals in 1424 families



The MitoPhen Database 1.7



Ratnaike, Greene et al. (2021), *Nucleic Acids Research*.

[Download database](#)

Select patients with variant

Select patients by PubMed ID

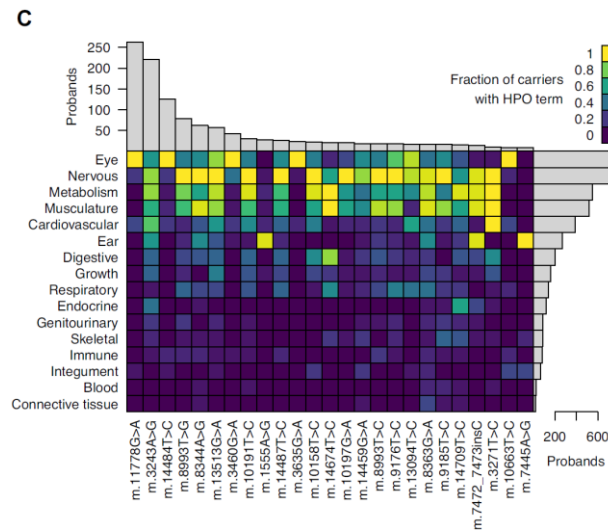
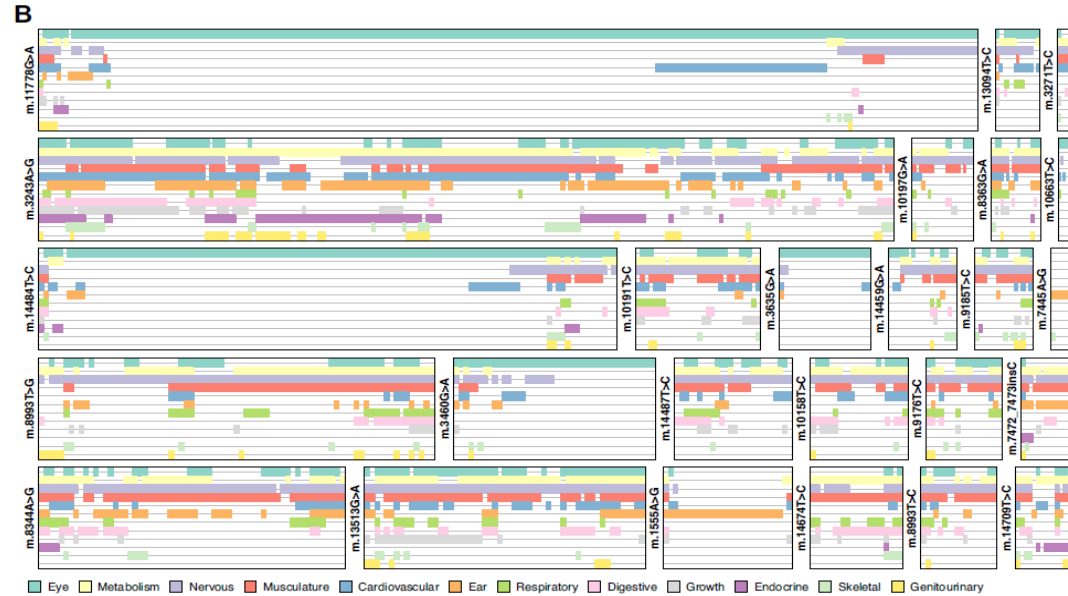
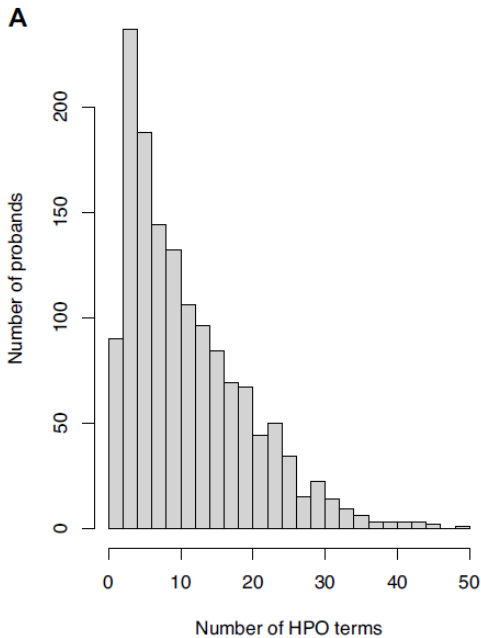
Select patients with HPO terms:
No terms selected

www.mitophen.org

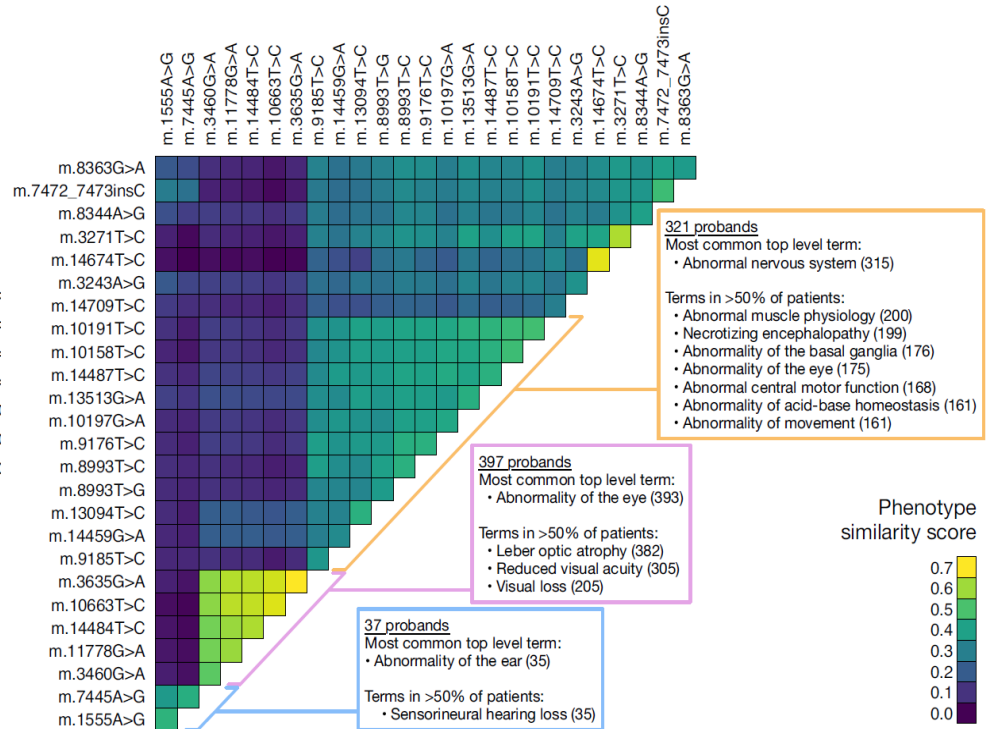
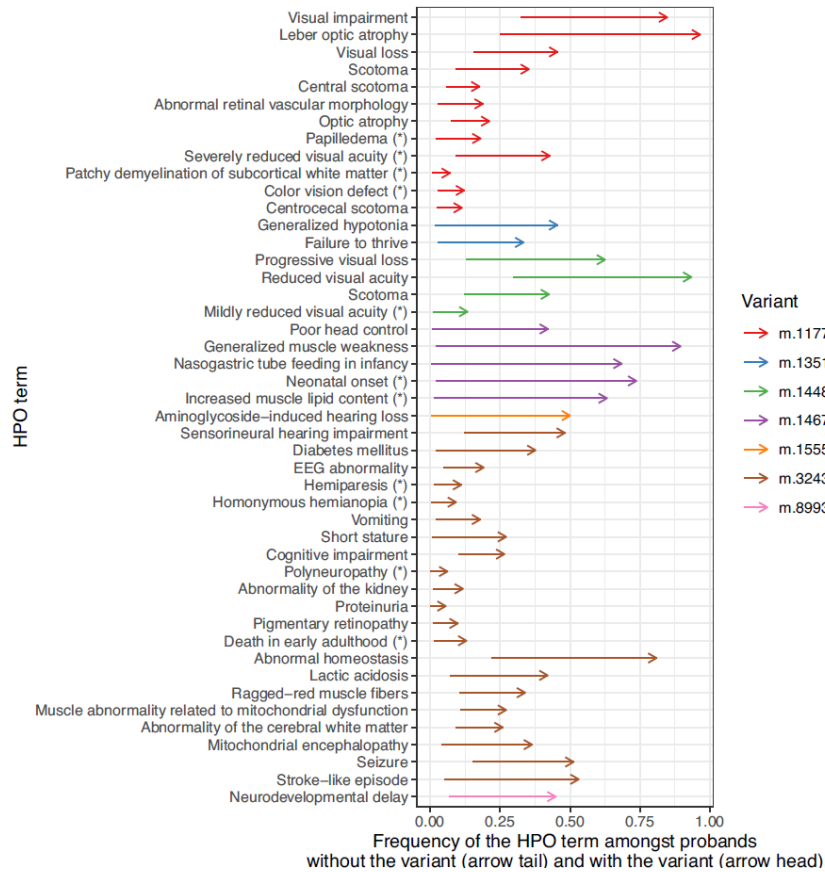
Refined phenotypes



- The **mean number of terms** per proband was **11.4**
- HPO terms:** nervous system, musculature, metabolism, cardiovascular, ear and eye



Refined phenotypes



New Phenotypes

LIG3



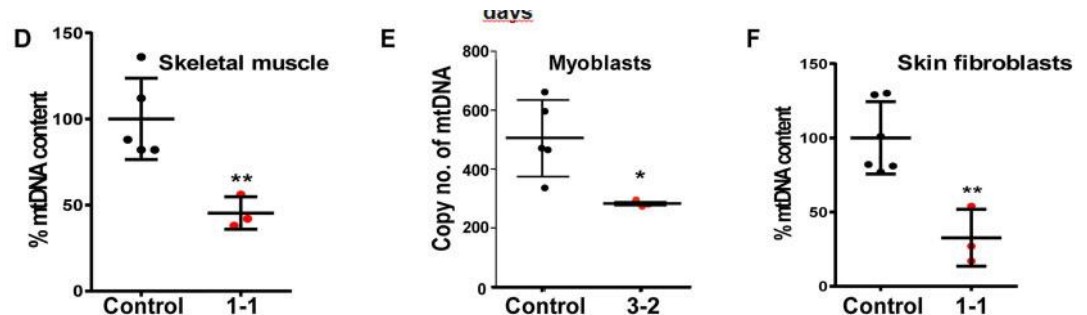
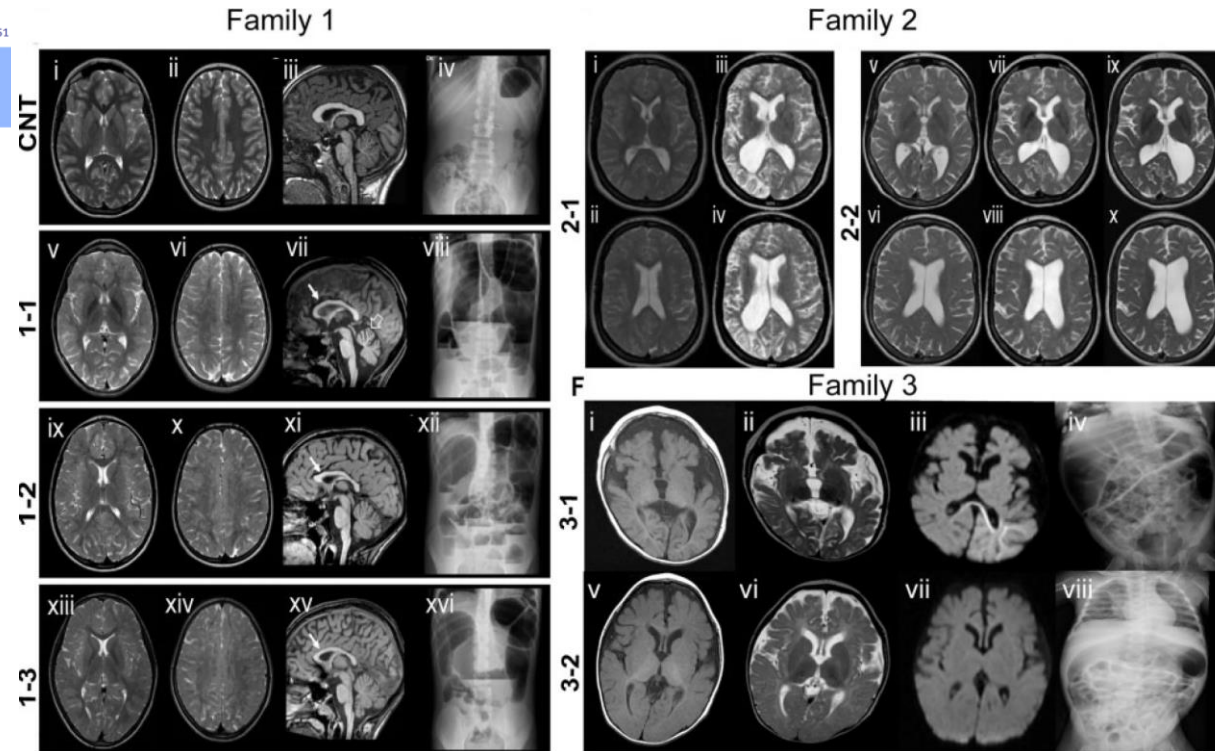
doi:10.1093/brain/awab056

BRAIN 2021; 144; 1451–1466 | 1451



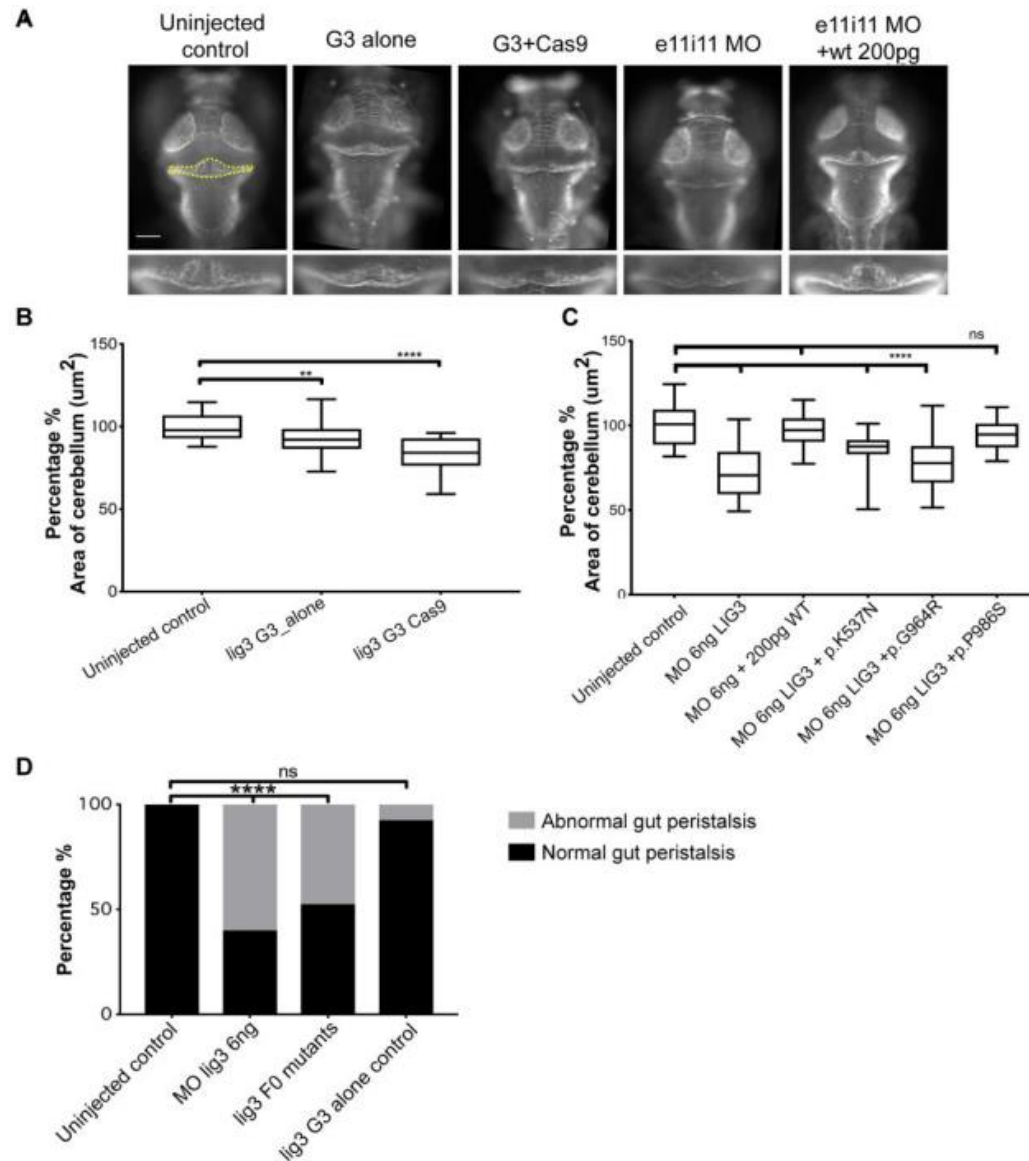
Biallelic variants in *LIG3* cause a novel mitochondrial neurogastrointestinal encephalomyopathy

- *LIG3* is the only ligase responsible for mitochondrial DNA (mtDNA) replication and maintenance
- **Seven** affected individuals
- neurogastrointestinal encephalomyopathy characterized by CIPO, neurogenic bladder, myopathic changes, and neurological impairment with stroke-like episodes, epilepsy and leukoencephalopathy

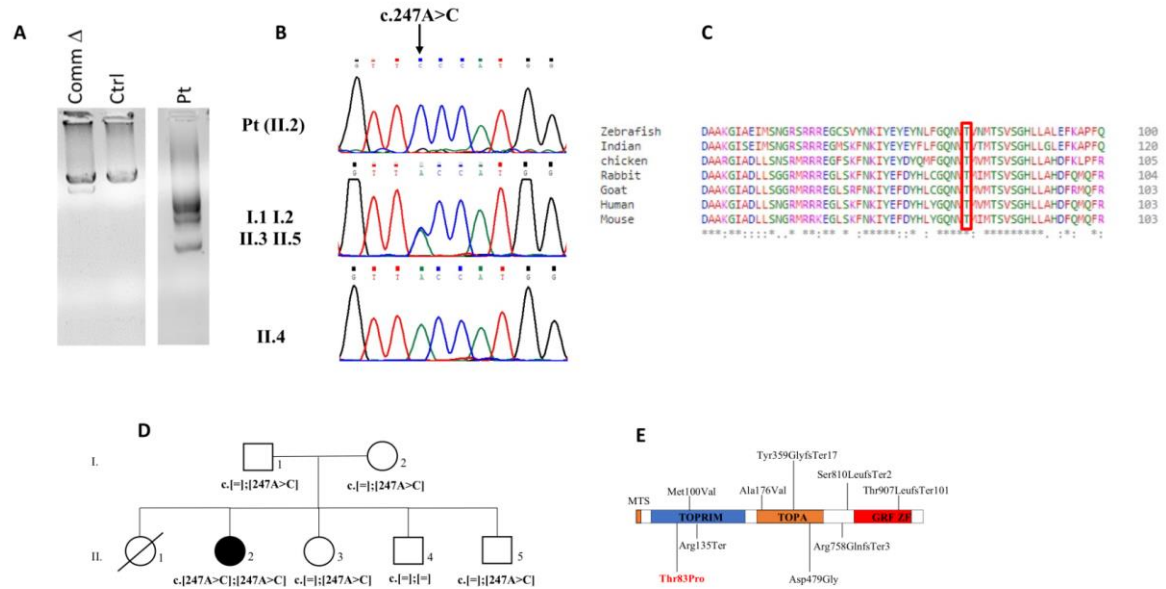
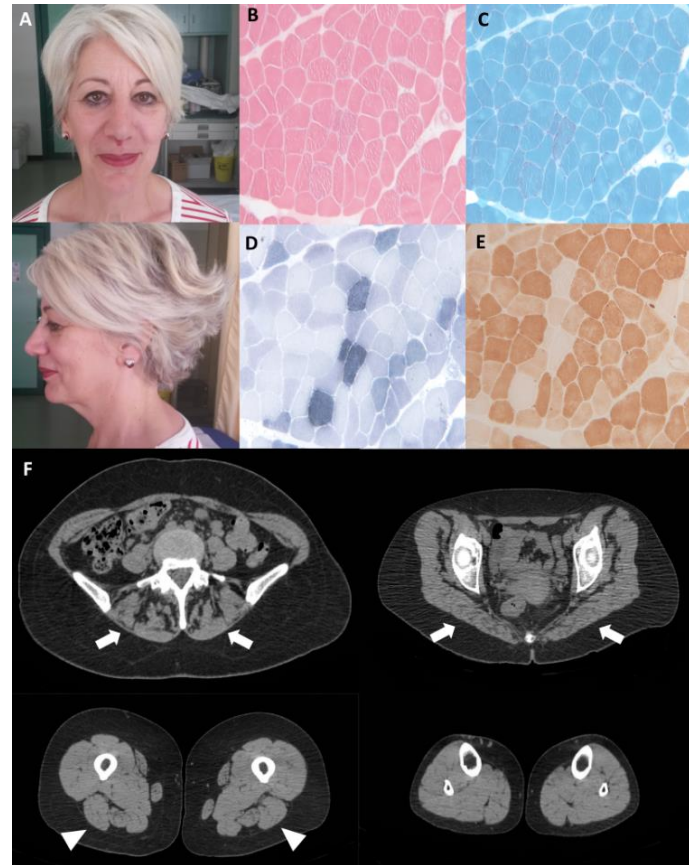


New Phenotypes

LIG3



New Phenotypes *TOP3A*

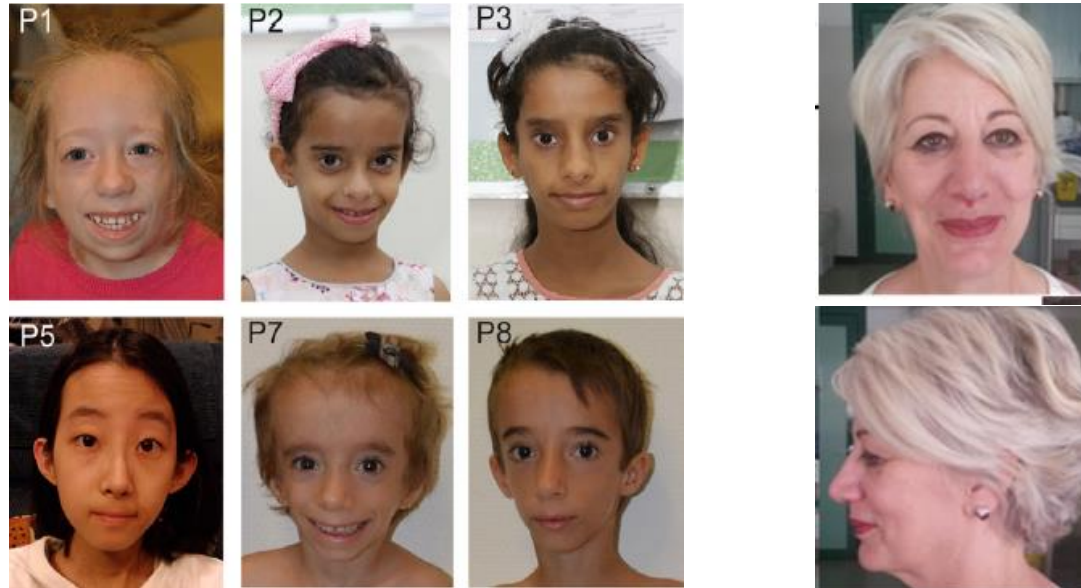


New Phenotypes

TOP3A

The American Journal of Human Genetics 103, 221–231, August 2, 2018 **ARTICLE**

Mutations in *TOP3A* Cause a Bloom Syndrome-like Disorder



Bloom syndrome-like disorder

PMM/PEO Plus

mtDNA depletion

Multiple mtDNA deletions

- **Similarities:** short stature, dilated cardiomyopathy, narrow face with prominent nose for paucity of subcutaneous fat, absence of malignancies
- **Differences:** a late onset of disease, no café'-au-lait macules

Expanding phenotypes

Movement disorders



Journal of Neurology
<https://doi.org/10.1007/s00415-021-10697-1>

ORIGINAL COMMUNICATION



Adult-onset mitochondrial movement disorders: a national picture from the Italian Network

V. Montano¹ · D. Orsucci² · V. Carelli^{3,4,5} · C. La Morgia^{3,4} · M. L. Valentino^{3,4,5} · C. Lamperti⁶ · S. Marchet⁶ · O. Musumeci⁷ · A. Toscano⁷ · G. Primiano^{8,9} · F. M. Santorelli¹⁰ · C. Ticci¹⁰ · M. Filosto¹¹ · A. Rubegni¹⁰ · T. Mongini¹² · P. Tonin¹³ · S. Servidei^{8,9} · R. Ceravolo¹ · G. Siciliano¹ · Michelangelo Mancuso¹⁰

Journal of
Clinical Medicine



Movement Disorders in Children with a Mitochondrial Disease: A Cross-Sectional Survey from the Nationwide Italian Collaborative Network of Mitochondrial Diseases

Chiara Ticci¹, Daniele Orsucci², Anna Ardisson³, Luca Bello⁴, Enrico Bertini⁵, Irene Bonato⁶, Claudio Bruno⁶, Valerio Carelli^{7,8}, Daria Diodato⁵, Stefano Doccini¹⁰, Maria Alice Donati⁹, Claudia Dosi¹, Massimiliano Filosto¹⁰, Chiara Fiorillo¹¹, Chiara La Morgia^{7,12}, Costanza Lamperti¹³, Silvia Marchet¹³, Diego Martinelli⁵, Carlo Minetti¹¹, Maurizio Moggio¹⁴, Tiziana Enrica Mongini¹⁵, Vincenzo Montano¹⁶, Isabella Moroni³, Olimpia Musumeci¹⁷, Elia Pancheri¹⁸, Elena Pegoraro⁴, Guido Primiano^{19,20}, Elena Procopio⁹, Anna Rubegni¹, Roberta Scalise^{1,21}, Monica Sciacco¹⁴, Serenella Servidei^{19,20}, Gabriele Siciliano¹⁶, Costanza Simoncini¹⁶, Deborah Tolomeo¹, Paola Tonin¹⁸, Antonio Toscano¹⁷, Flavia Tubili⁹, Michelangelo Mancuso¹⁶, Roberta Battini^{1,16,*} and Filippo Maria Santorelli¹⁰



POLG c.428C>T [Ala143Val], 2956T>G [Tyr986Asp]

Expanding phenotypes

Spinal cord involvement



Communication

Spinal Cord Involvement in Adult Mitochondrial Diseases: A Cohort Study

Guido Primiano^{1,2,*}, Paolo Mariotti¹, Ida Turrini¹, Cristina Sancricca¹, Andrea Sabino², Alessandra Torracco³, Rosalba Carrozzo³ and Serenella Servidei^{1,2}

- 9,8%, 5/51 patients

Neurological Sciences (2022) 43:2081–2084
<https://doi.org/10.1007/s10072-022-05881-8>

BRIEF COMMUNICATION



Kearns-Sayre syndrome: expanding spectrum of a “novel” mitochondrial leukomyeloencephalopathy

Marco Moscatelli¹ · Anna Ardissoni² · Eleonora Lamantea³ · Giovanna Zorzi² · Claudio Bruno⁴ · Isabella Moroni² · Alessandra Erbetta¹ · Luisa Chiapparini¹

Neuroradiology (2020) 62:1315–1321
<https://doi.org/10.1007/s00234-020-02501-0>

PAEDIATRIC NEURORADIOLOGY



Spinal cord involvement in Kearns-Sayre syndrome: a neuroimaging study

Pasquini Luca^{1,2} · Guamera Alessia^{1,2} · Rossi-Espagnet Maria Camilla^{1,2} · Napolitano Antonio³ · Martinelli Diego⁴ · Deodato Federica⁴ · Diodato Daria⁵ · Carrozzo Rosalba⁵ · Dionisi-Vici Carlo⁴ · Longo Daniela¹

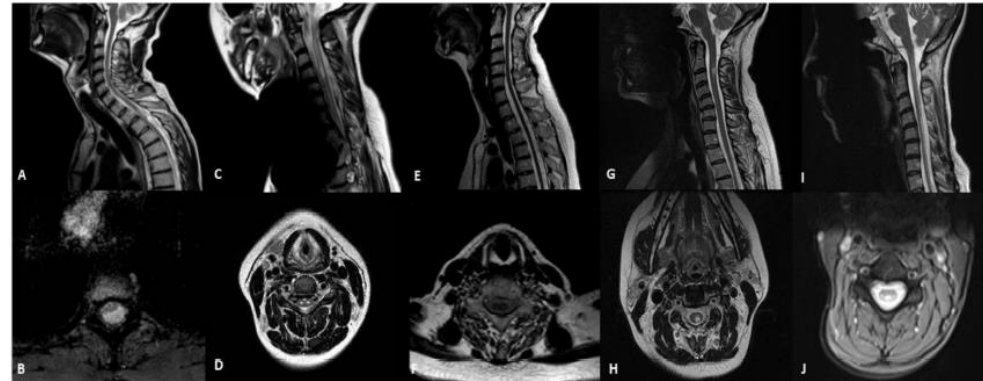
- 54.5%, 6/11

AJNR Am J Neuroradiol 42:389–96 Feb 2021

Involvement of the Spinal Cord in Primary Mitochondrial Disorders: A Neuroimaging Mimicker of Inflammation and Ischemia in Children

C.A.P.F. Alves, A. Goldstein, S.R. Teixeira, J.S. Martin-Saavedra, I.P. de Barcelos, G. Fadda, L. Caschera, M. Kidd, F.G. Gonçalves, E.M. McCormick, M.J. Falk, Z. Zolkipli-Cunningham, A. Vossough, and G. Zuccoli

- 58%, 19/33



Sleep-Disordered Breathing in Adult Patients With Mitochondrial Diseases

A Cohort Study

Guido Primiano, MD, Valerio Brunetti, MD, Catello Vollono, MD, PhD, Anna Losurdo, MD, Rossana Moroni, PhD, Giacomo Della Marca, MD, and Serenella Servidei, MD

Neurology® 2021;96:e241–e249. doi:10.1212/WNL.00000000000011005

Correspondence

Dr. Primiano

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Figure 1 Distribution of the SDB in Mitochondrial Diseases

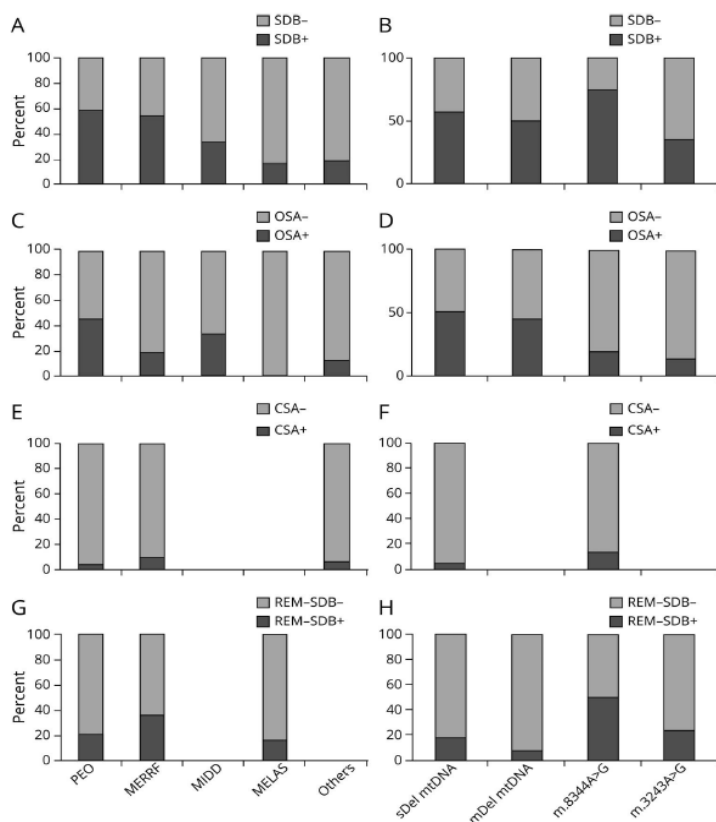
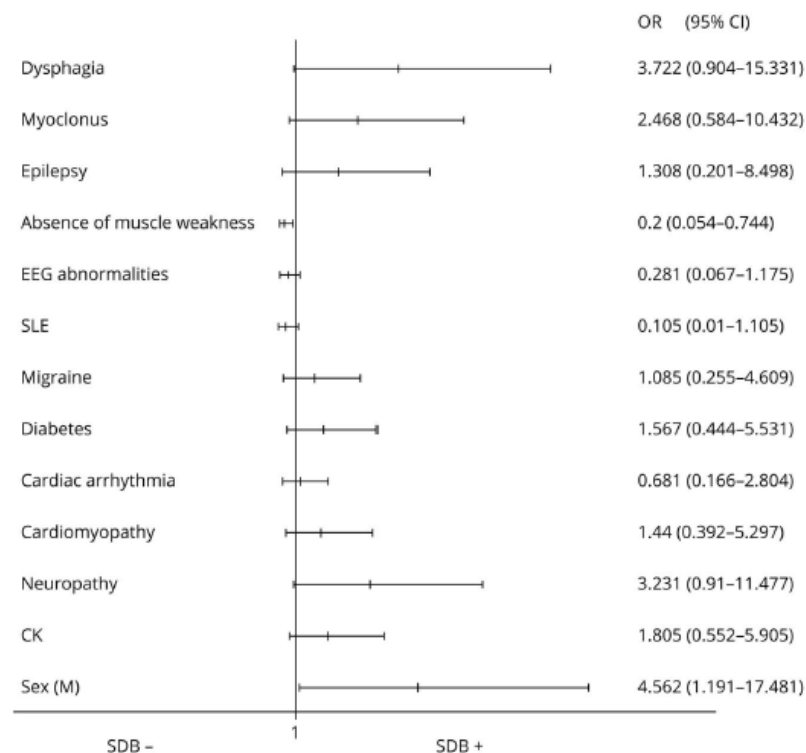


Figure 2 Association Between SDB and Clinical Variables





Mitochondrial disease and endocrine dysfunction

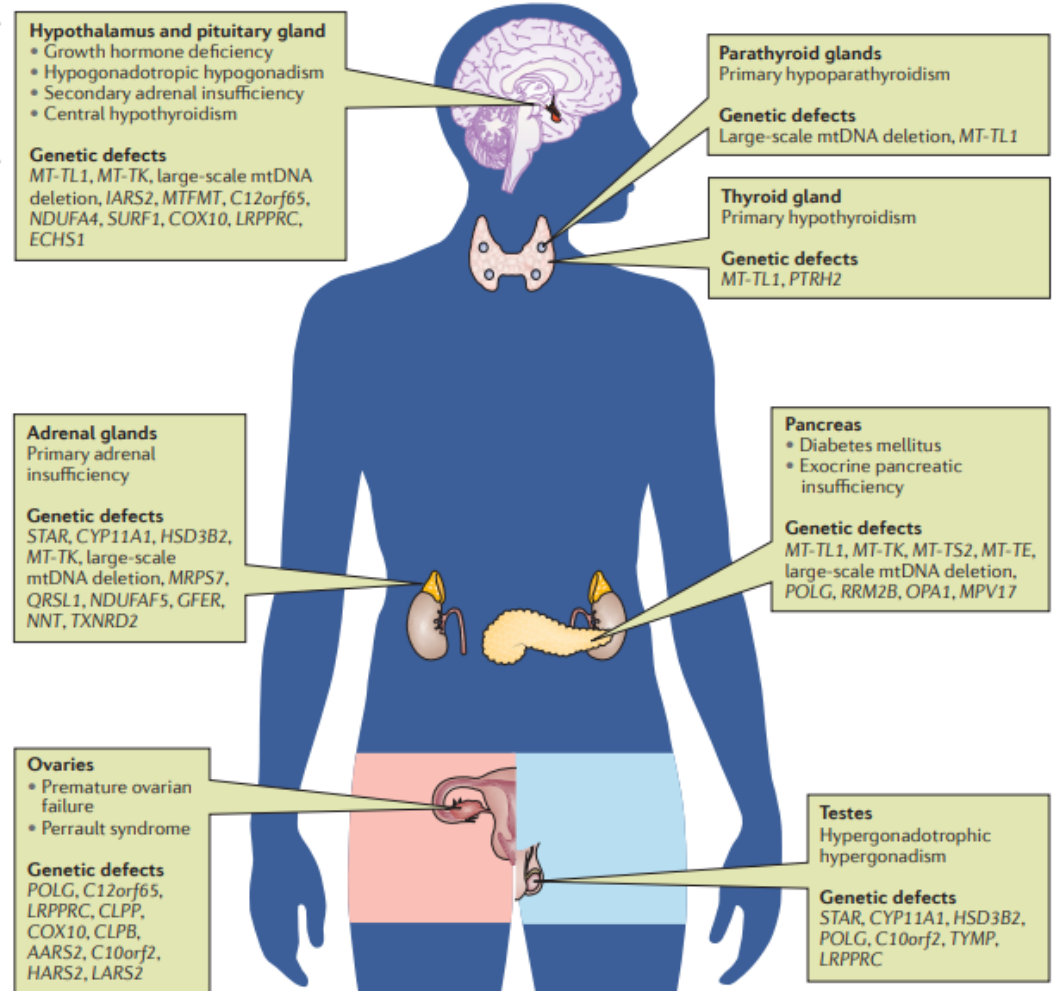
Jasmine Chow¹, Joyeeta Rahman², John C. Achermann², Mehul T. Dattani^{2,3} and Shamima Rahman^{2,4}



Mitochondrial Dysfunction in Primary Ovarian Insufficiency


Dov Tiosano, Jason A. Mears, and David A. Buchner

Endocrinology
Endocrine Society



Safety of drug use in patients with a primary mitochondrial disease: An international Delphi-based consensus

A severe linezolid-induced rhabdomyolysis and lactic acidosis in Leigh syndrome

Guido Primiano¹ 
Serenella Servidei^{1,2}


Maaïke C. De Vries¹  | David A. Brown² | Mitchell E. Allen² |
Laurence Bindoff^{3,4} | Gráinne S. Gorman^{5,6} | Amel Karaa⁷ |
Nandaki Keshavan^{8,9} | Costanza Lamperti¹⁰ | Robert McFarland^{5,6} |
Yi Shiao Ng^{5,6} | Mar O'Callaghan^{11,12} | Robert D. S. Pitceathly¹³ |
Shamima Rahman^{8,9} | Frans G. M. Russel¹⁴ | Kristin N. Varhaug^{3,4} |
Tom J. J. Schirris¹⁴ | Michelangelo Mancuso¹⁵

TABLE 2 Points of attention regarding drug prescription in patients with a mitochondrial disease (detailed description in Section 4)

Specific drug/drug group/clinical condition/genotype	Points of attention
<i>Specific drug/drug group/genotype</i>	
Aminoglycosides	The mitochondrial 12S rRNA is a hot spot for mutations associated with both aminoglycoside-induced and non-syndromic hearing loss. Screening for these mtDNA mutations is strongly recommended before elective long-term treatment is planned. The benefits of the drug in emergency treatment, as a very effective broad-spectrum antibiotic, outweigh the risks in these situations.
Valproic acid	Should be used only in exceptional circumstances. The drug is absolutely contraindicated in patients with mitochondrial disease due to <i>POLG</i> mutations. Valproic acid should not be used in patients with known liver disease and/or clinical signs suspicious for <i>POLG</i> disease.
Neuromuscular blocking agents	Extra caution and monitoring should be performed for patients manifesting a predominantly myopathic phenotype.
<i>Specific clinical condition</i>	
General anaesthesia and surgery	Catabolism should be prevented by minimising preoperative fasting and administering intravenous glucose perioperatively during prolonged anaesthesia, unless the patient is on a ketogenic diet.
Duration of treatment	The duration of drug administration may play a role in whether or not side effects develop. Duration of treatment should be guided by individual patient needs and their response to specific treatments.
Renal impairment	Many patients with a mitochondrial disease have renal impairment; drug dose adjustment should be considered particularly when active drug moieties are renally cleared.
Metabolic acidosis (lactic acidosis)	Metabolic acidosis (lactic acidosis) may occur in patients with mitochondrial disease, therefore drugs that can cause acidosis should be prescribed with caution. Regular clinical review and monitoring of acid-base status in blood is recommended.



Mitochondrion 58 (2021) 243–245

SARS-CoV-2 infection in patients with primary mitochondrial diseases: Features and outcomes in Italy

Michelangelo Mancuso^{a,*}, Chiara La Morgia^b, Maria Lucia Valentino^{b,c}, Anna Ardisson^d,
Costanza Lamperti^e, Elena Procopio^f, Caterina Garone^{g,h}, Gabriele Siciliano^a,
Olimpia Musumeciⁱ, Antonio Toscanoⁱ, Guido Primiano^{j,k}, Serenella Servidei^{j,k},
Valerio Carelli^{b,c}

CLINICAL/SCIENTIFIC NOTE

OPEN ACCESS

COVID-19–Related Outcomes in Primary Mitochondrial Diseases

An International Study

Chiara Pizzamiglio, MD,* Pedro M. Machado, MD, PhD,* Rhys H. Thomas, MD, PhD,
Gráinne S. Gorman, MD, PhD, Robert McFarland, MD, PhD, Michael G. Hanna, MD, FRCP, and
Robert D. S. Pitceathly, MD, PhD, on behalf of the MitoCOVID-19 Study Group

Neurology® 2022;98:576–582. doi:10.1212/WNL.0000000000200240

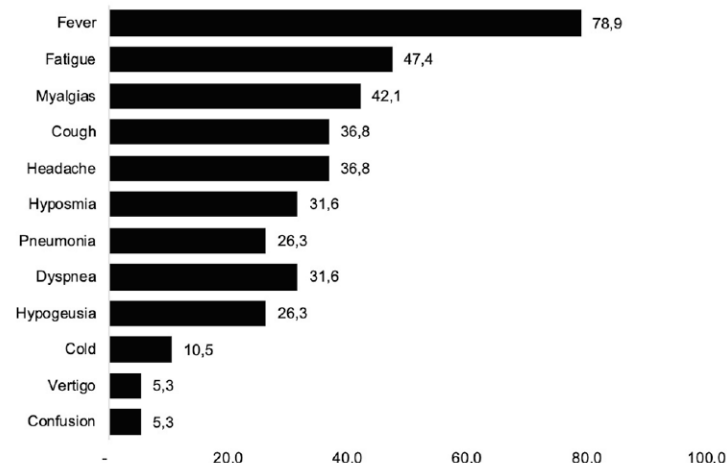


Fig. 1. Percentage of COVID-19-related symptoms in the 27 symptomatic mitochondrial patients.

- **Seventy-nine PMDs** from 10 countries (mean age 41.5 ± 18 years);
- 25 (32%) were **hospitalized**, 48 (61%) recovered fully, 28 (35%) improved with sequelae, and 3 (4%) **died**;
- Statistically significant differences in **hospitalization status** were observed in baseline status, including the **NMDAS score** ($p = 0.003$) and mRS ($p = 0.001$), presence of **respiratory dysfunction** ($p < 0.001$), **neurologic involvement** ($p = 0.003$), and **more than 4 comorbidities** ($p = 0.002$).
- **Respiratory dysfunction** is an independent risk factor for severe COVID-19 in PMDs while high **disease burden** and **coexisting comorbidities** contribute toward COVID-19–related hospitalization.



Neurodegenerative and functional signatures of the cerebellar cortex in m.3243A > G patients

©Roy A. M. Haast,¹ Irena F. M. De Co,² Dimo Ivanov,³ Ali R. Khan,^{1,4,5} Jacobus F. A. Jansen,^{6,7,8} Hubert J. M. Smeets^{2,8} and Kamil Uludag^{9,10,11}

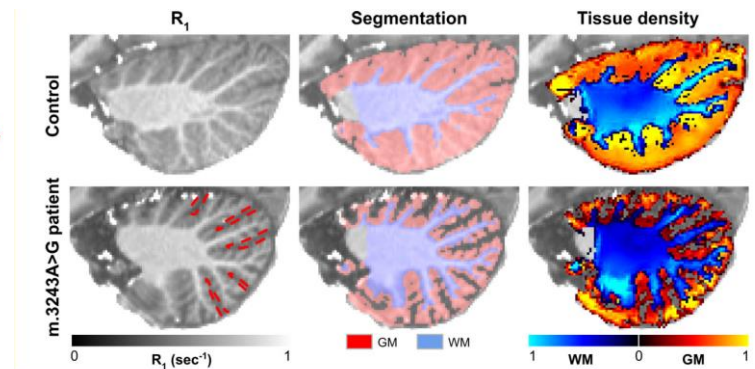
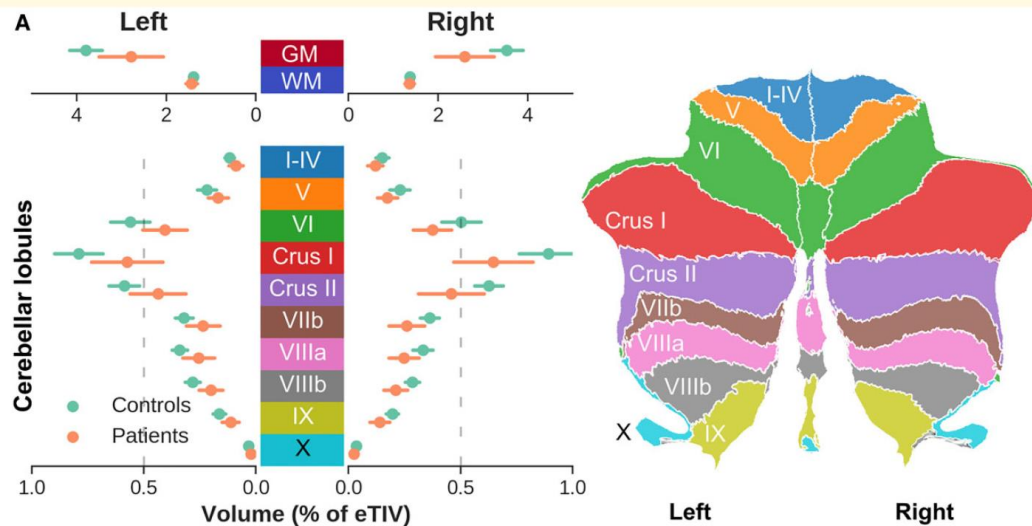


Figure 1 Example data. Left to right: R_1 , GM (red) and WM (blue) segmentation masks and corresponding tissue density maps are shown for a control (top row) and m.3243A > G patient (bottom row). Dashed red lines indicate the inter-folial spacing for the patient.

Figure 2 Cerebellar GM and WM volumes. **(A)** Comparison of volume (presented as % of eTIV) on the x-axis between controls (green) and m.3243A > G patients (orange) for left and right hemisphere GM and WM (top), as well as per cerebellar lobule GM (bottom), colour-coded based on the right panel legend. **(B)** First two columns: correlation between GM volume (y-axis) and NMDAS or corrected UEC mutation load (x-axes). Last two columns: similar to first two columns but using WM volume (y-axis). Shaded areas show 95% confidence intervals.

Mitochondrial diseases



Mitochondrial Diseases

Gemelli

Fondazione Policlinico Universitario A. Gemelli
Università Cattolica del Sacro Cuore



Medicina mitocondriale



Gemelli

Fondazione Policlinico Universitario Agostino Gemelli IRCCS
Università Cattolica del Sacro Cuore

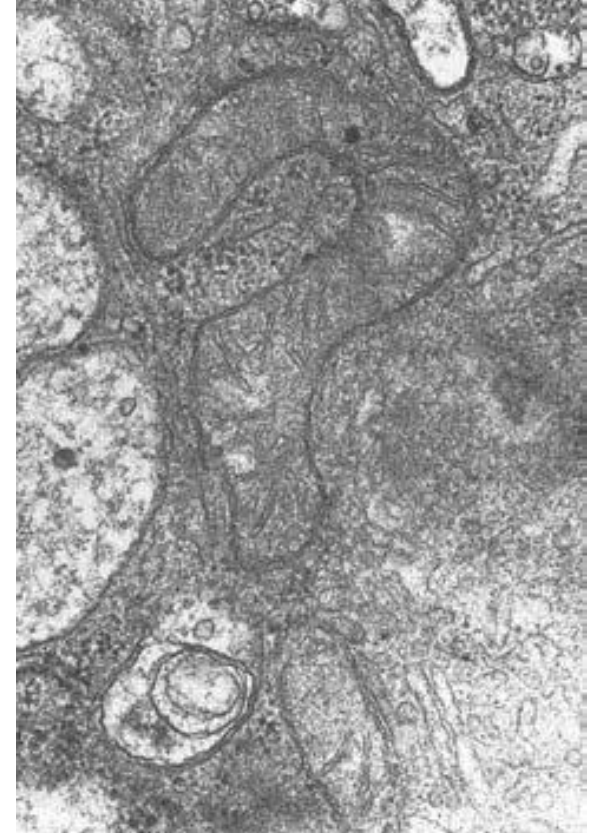


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